

# MOLECULAR ANALYSIS OF ACUTE PROMYELOCYTIC LEUKEMIA BY NEXT GENERATION SEQUENCING

## 7th INTERNATIONAL SYMPOSIUM ON ACUTE PROMYELOCYTIC LEUKEMIA

Llop M(1,9), Gil JV(2), Sargas C(2), Cervera J(3,9), Such E(3,9), Gil C(4), Sayas MJ(5), García R(6), Manso F(7), Fernández JM(8), Martínez-Cuadrón D(3), Rodríguez R(3), Boluda B(3), Montesinos P(3), Sanz MA(3,9) Barragán E(1,9).

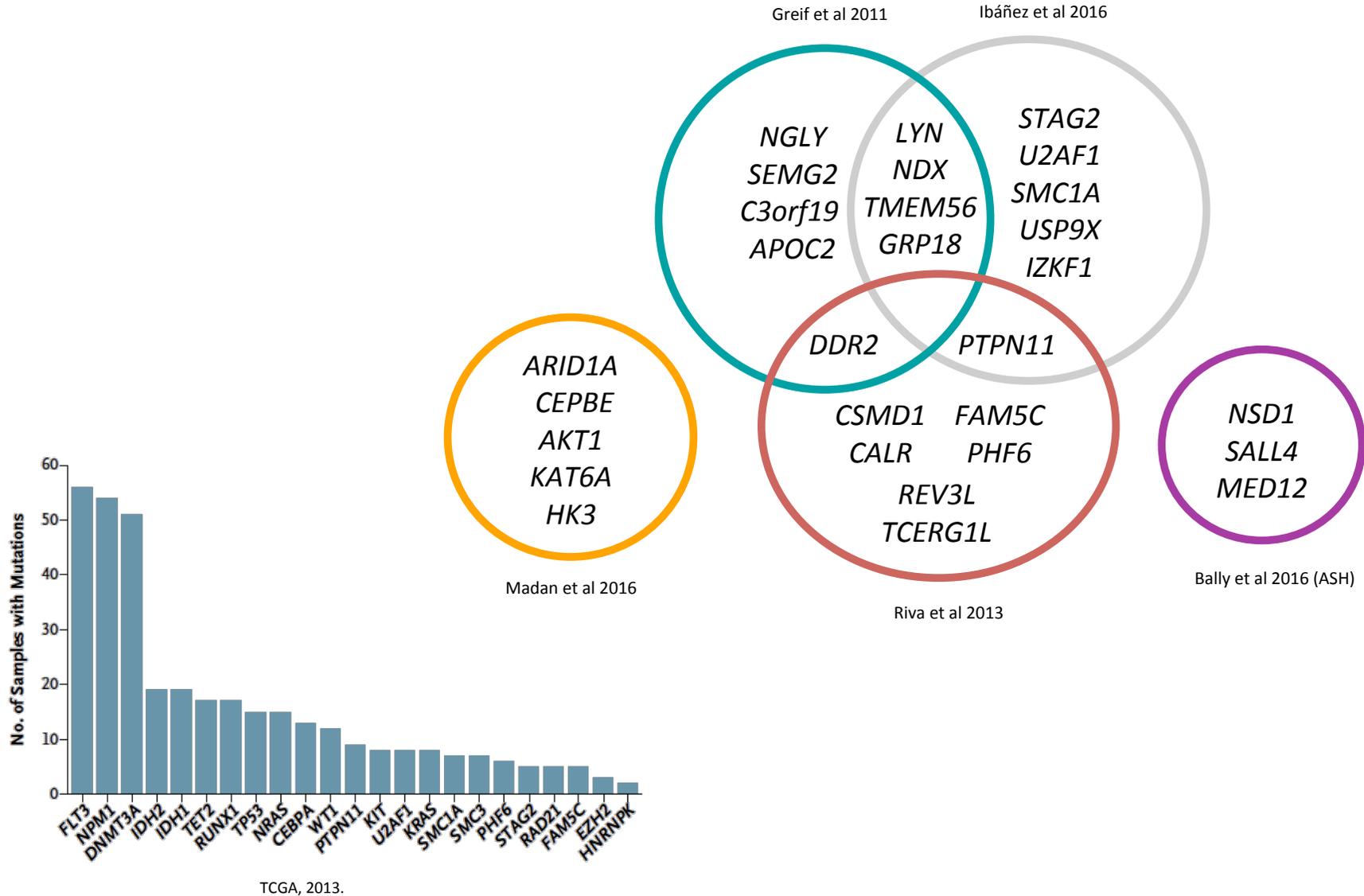


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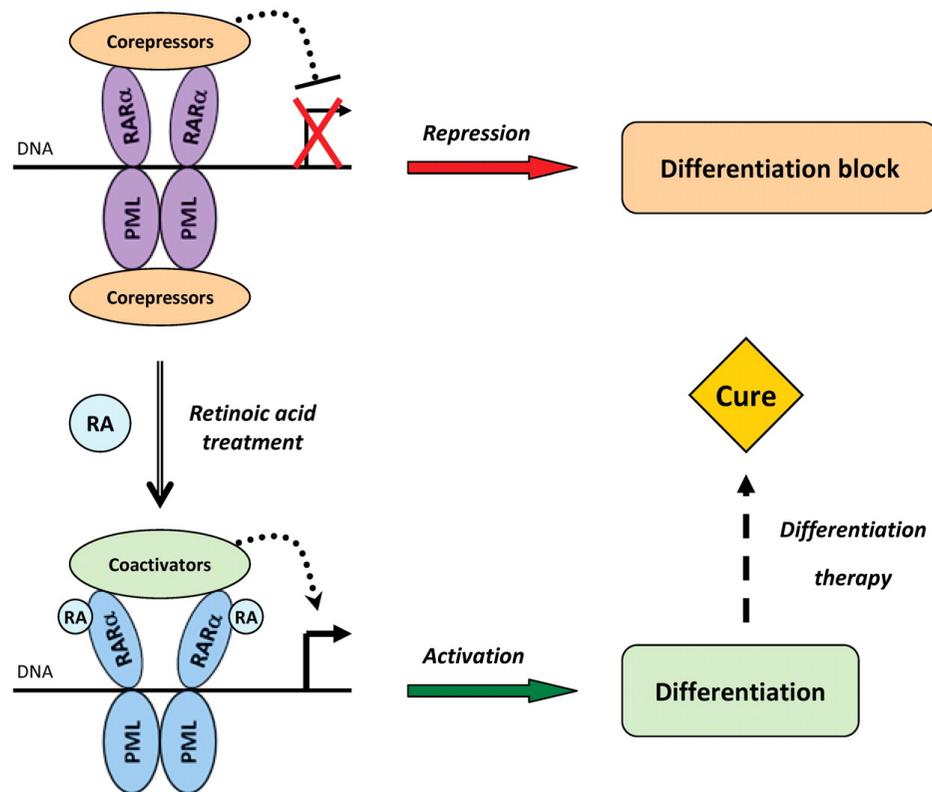
# Introduction

## Revealing the genetic landscape of AML



# Introduction

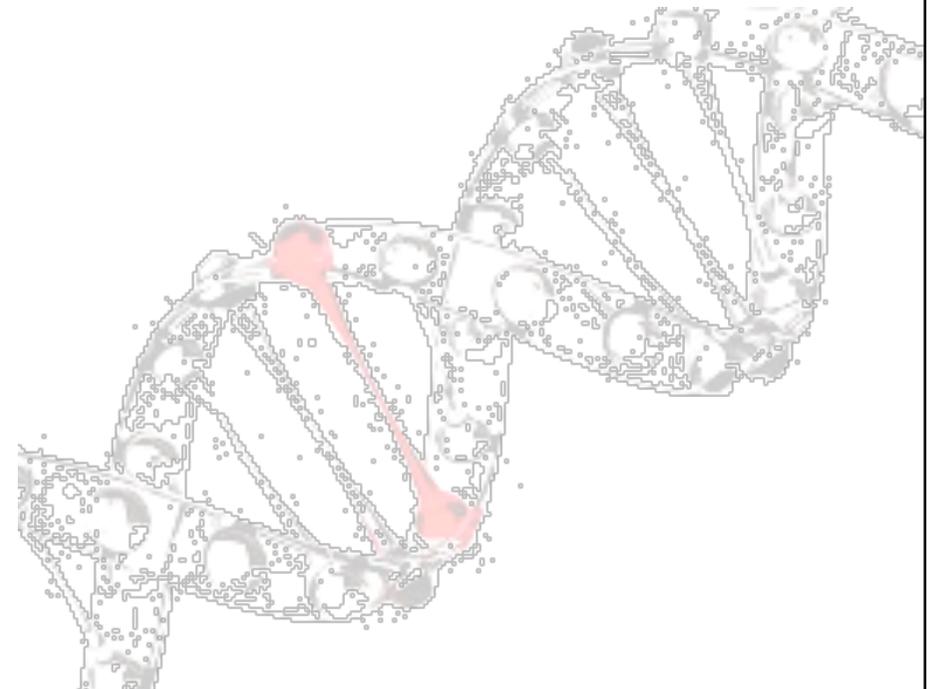
## Revealing the genetic landscape of AML



Ablain and de The 2011

# Aims

The aim of this study is to analyze the mutational landscape of favourable AML with NGS, focusing on APL

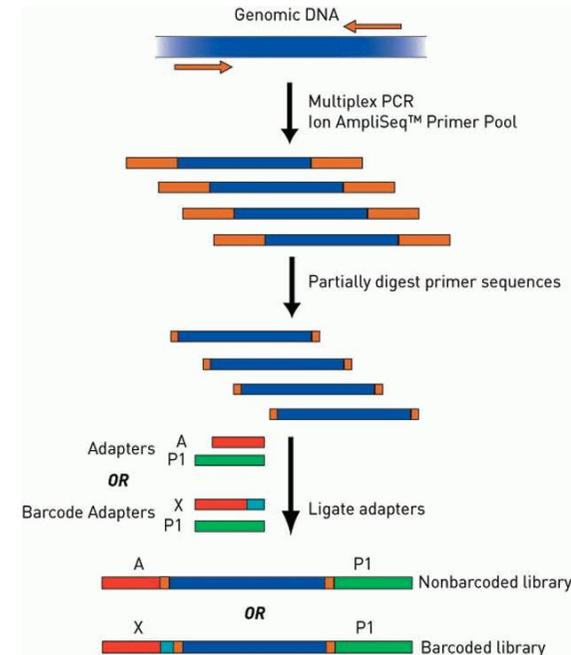


# Patients and Methods

## Next generation sequencing

### AML AMPLISEQ COMMUNITY PANEL

Gen	Chr.	Amplicones	Bases	CDS exons
<i>ASXL1</i>	chr20	28	2947	12
<i>BRAF</i>	chr7	1	11	15 (V600E)
<i>CBL</i>	chr11	5	416	8, 9
<i>CEBPA</i>	chr19	9	1117	CDS
<i>DNMT3A</i>	chr2	42	3619	CDS
<i>FLT3</i>	chr13	3	66	16 (N676), 20-21 (830-850)
<i>GATA2</i>	chr3	20	1643	CDS
<i>IDH1</i>	chr2	3	332	4
<i>IDH2</i>	chr15	2	201	4
<i>JAK2</i>	chr9	1	128	14
<i>KIT</i>	chr4	8	632	8, 10, 11, 17
<i>KRAS</i>	chr12	4	370	2, 3, 4
<i>NPM1</i>	chr5	1	79	11
<i>NRAS</i>	chr1	3	370	2, 3, 4
<i>PTPN11</i>	chr12	5	427	3, 7, 8, 13
<i>RUNX1</i>	chr21	15	1149	3-8
<i>TET2</i>	chr4	59	6369	CDS
<i>TP53</i>	chr17	24	1715	CDS
<i>WT1</i>	chr11	4	324	7,9



Ion Chef

Clonal amplification on ISPs by emPCR and template-positive ISP enrichment (+ chip load)



PGM

Sequencing by synthesis. Semiconductor technology

# Patients and Methods

## Favorable-risk AML

73 favorable risk AML

45 APL (61.64%)

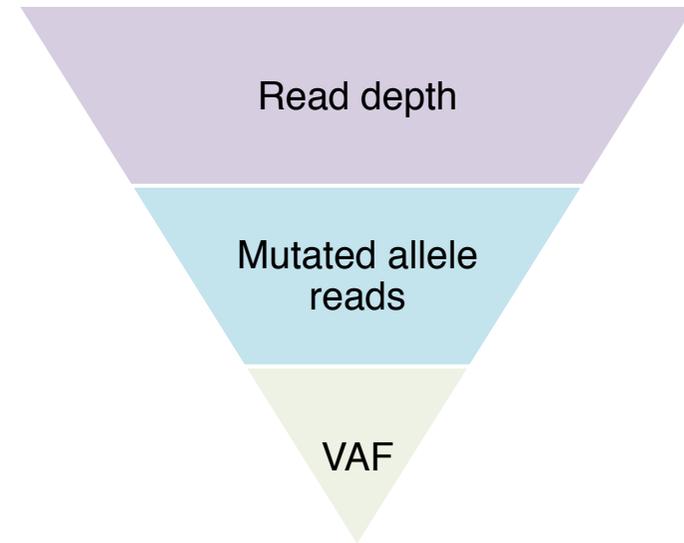
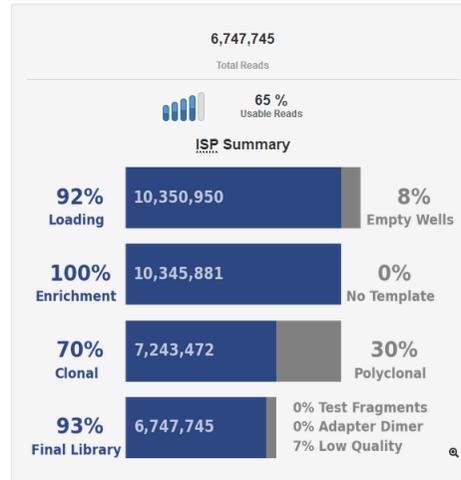
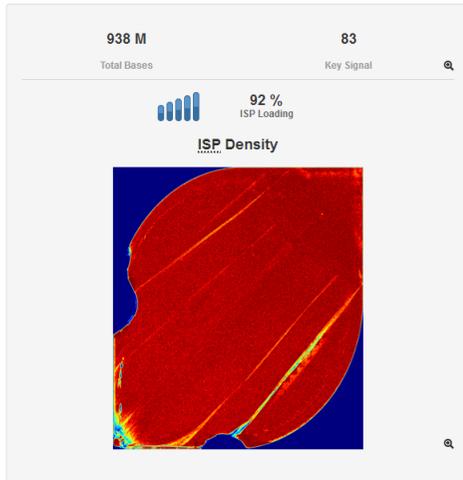
28 no promyelocytic AML (38.36%)

APL	N (%)	Median (range)
<b>Age</b>		47 (10-77)
Adult	39 (86.67)	
Pediatric	6 (13.33)	
<b>Sex</b>		
Male	18 (40)	
Female	27(60)	
<b>APL type</b>		
Primary	37 (82.22)	
Secondary	8 (17.78)	
<b>PML-RARA type</b>		
BCR1	29 (64.44%)	
BCR3	16 (35.56)	

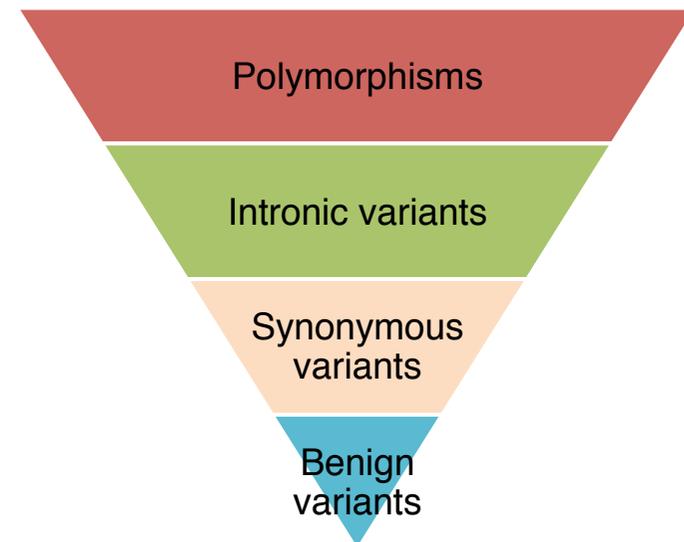
No APL AML	N (%)	Median (range)
<b>Age</b>		45.5 (19-67)
Adult	28 (100)	
Pediatric	0 (0)	
<b>Sex</b>		
Male	19(68)	
Female	9 (32)	
<b>Rearrangement</b>		
<i>AML1-ETO</i>	15 (53.57)	
<i>CBFB-MYH11</i>	13 (46.43)	

# Results

## Sequencing metrics and variant analysis



	MAPPED READS (bp)	ON TARGET (%)	MEAN DEPTH (reads)	UNIFORMITY (%)
Mean	820350	90.28	3231	93.19
Min	669428	56.56	1028	89.44
Max	986560	98.66	4760	96.50



Analysis Results

Analysis Name: 46143\_APL

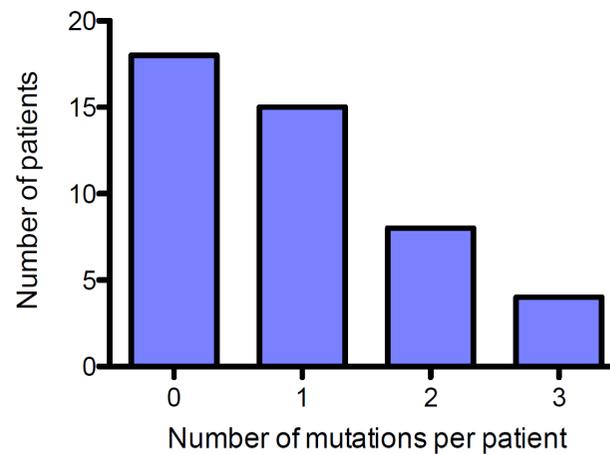
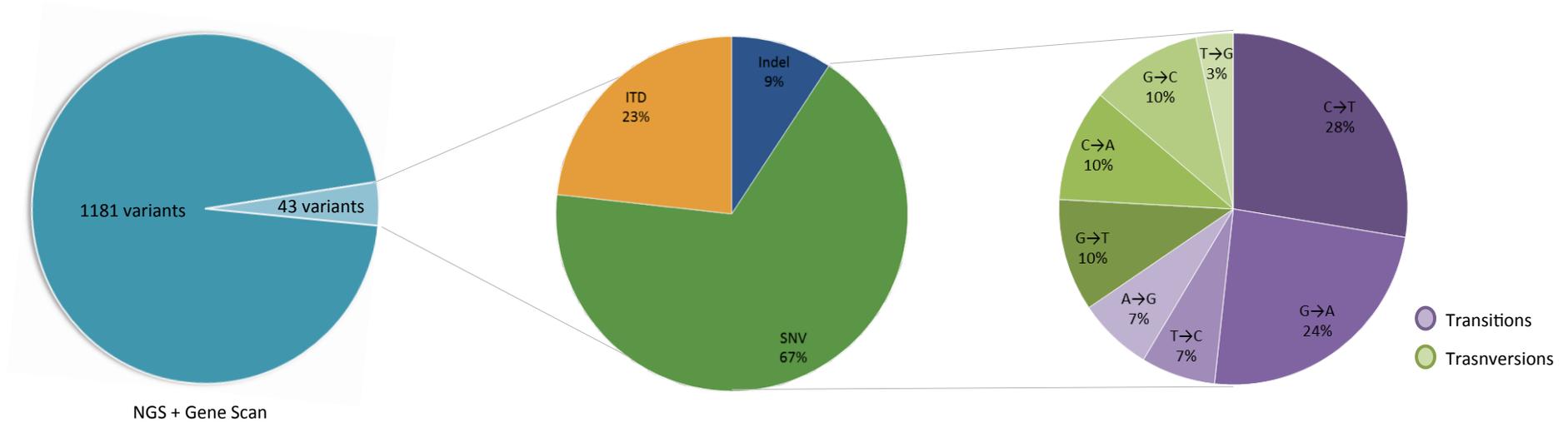
Summary Functional Population Ontologies Pharmacogenomics Somatic QC

...	Classification	Locus	Genotype	Ref	Type	No Call Reason	Genes	Location	Length	Variant ID
☐	Unclassified	chr4:106198248	GCCCAAGCCCA	GCCCCA	INDEL		TET2	exonic	1	
☐	Unclassified	chr11:32413566	G/A	G	SNV		WT1	exonic	1	
☐	Unclassified	chr11:32413578	G/A	G	SNV		WT1	exonic	1	
☐	Unclassified	chr11:32417947	G/A	G	SNV		WT1	exonic	1	
☐	Unclassified	chr13:28602368	CGT/TG	TGG	MNV/INDEL		FLT3	exonic, exonic	3	
☐	Unclassified	chr19:33792729	G/A	G	SNV		CEBPA	exonic	1	

1 - 6 of 6 items

# Results

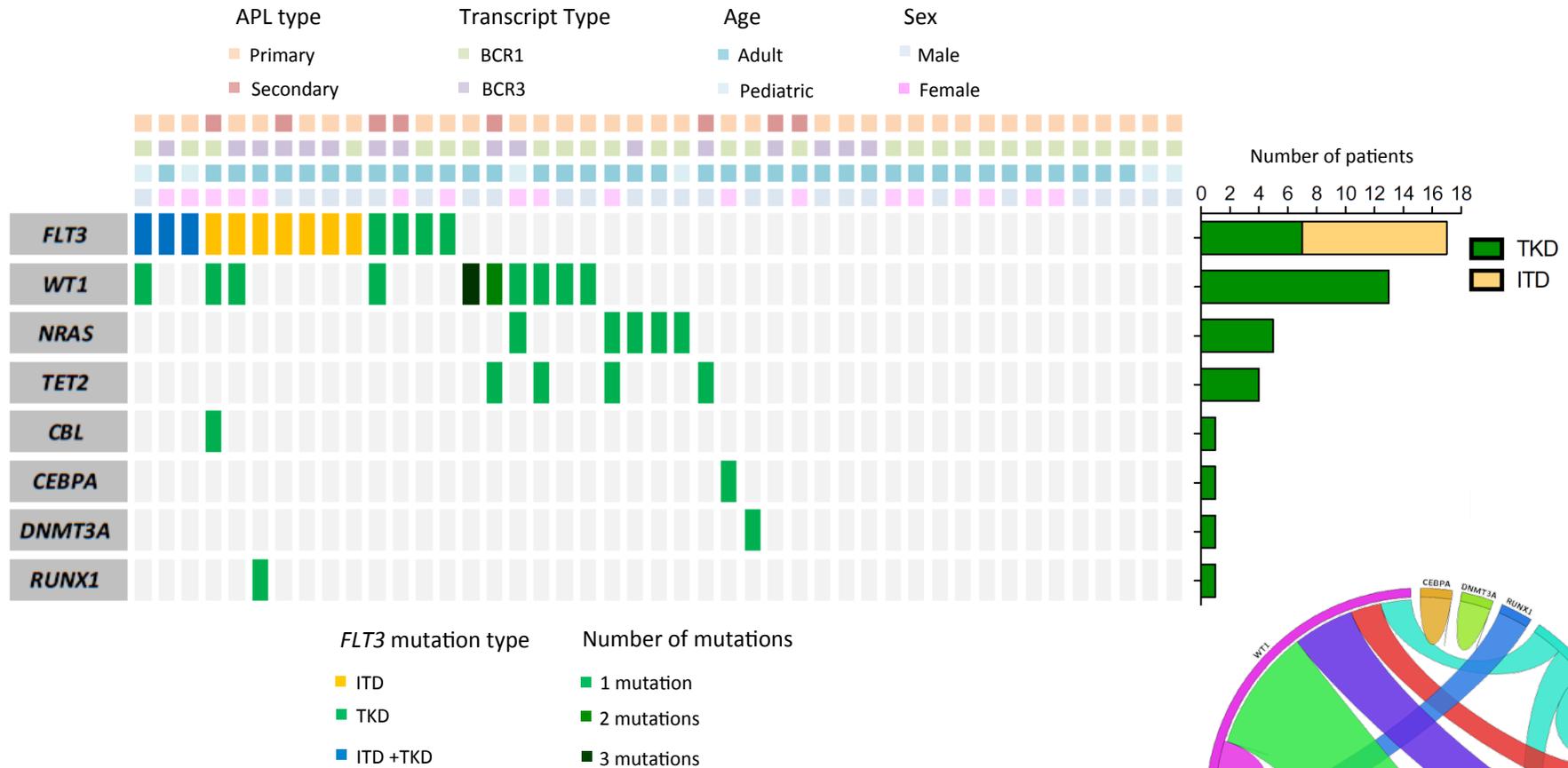
## Mutational distribution in APL



- 27/45 (60.0%) of patients showed at least one mutation
- The mean number of mutations per patient was 0.96

# Results

## Mutational distribution in APL

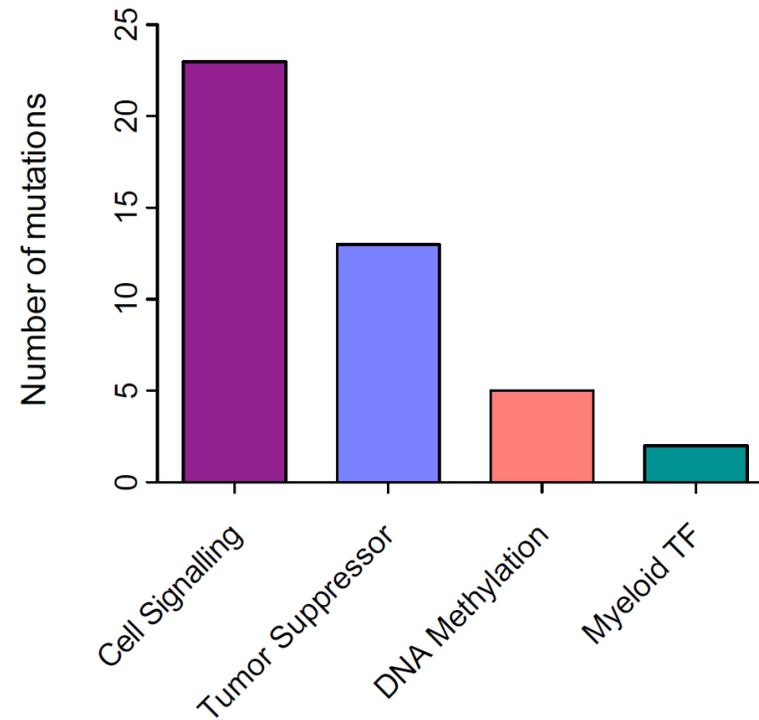
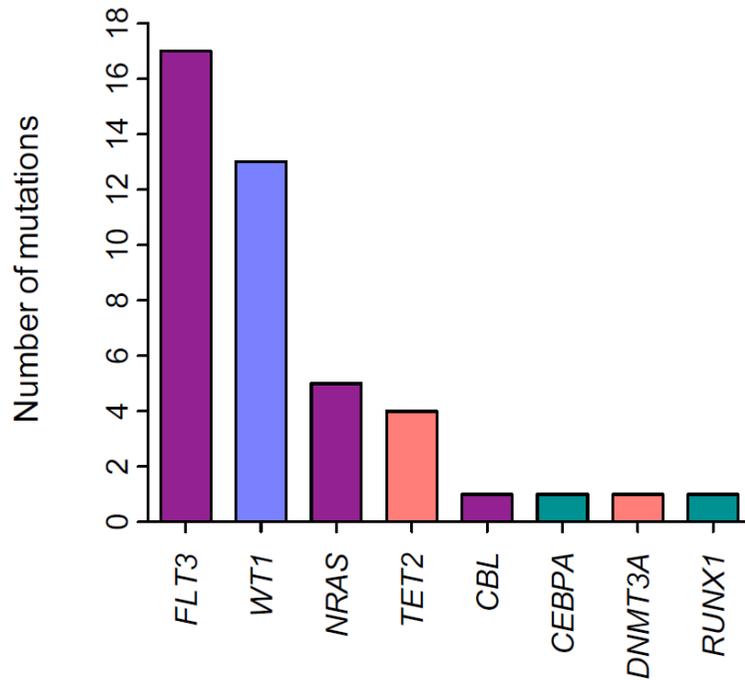


In 10/45 (22%) patients, *FLT3* was the only genetic mutation

Mutations in *RUNX1*, *DNMT3A*, *CEBPA* and *CBL* were found in less than 10% of cases

# Results

## Functional categories (TCGA)

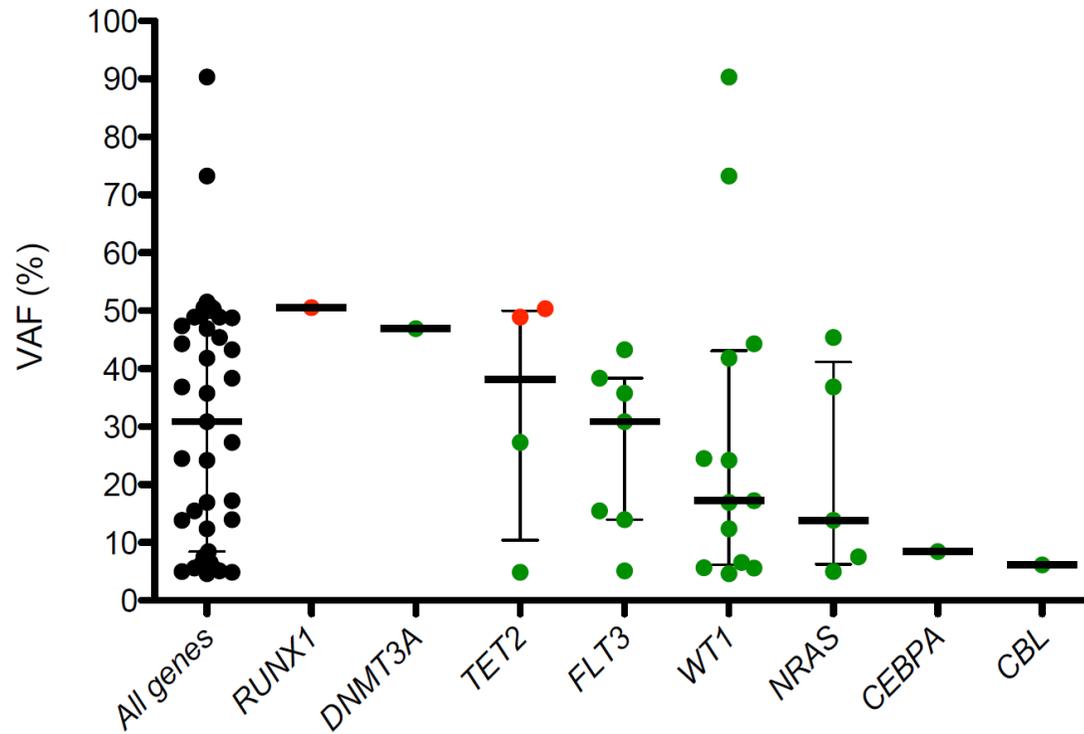


● Cell signalling    ● Tumor Suppressor    ● DNA methylation    ● Myeloid Transcription Factor

Transcription factor fusions

# Results

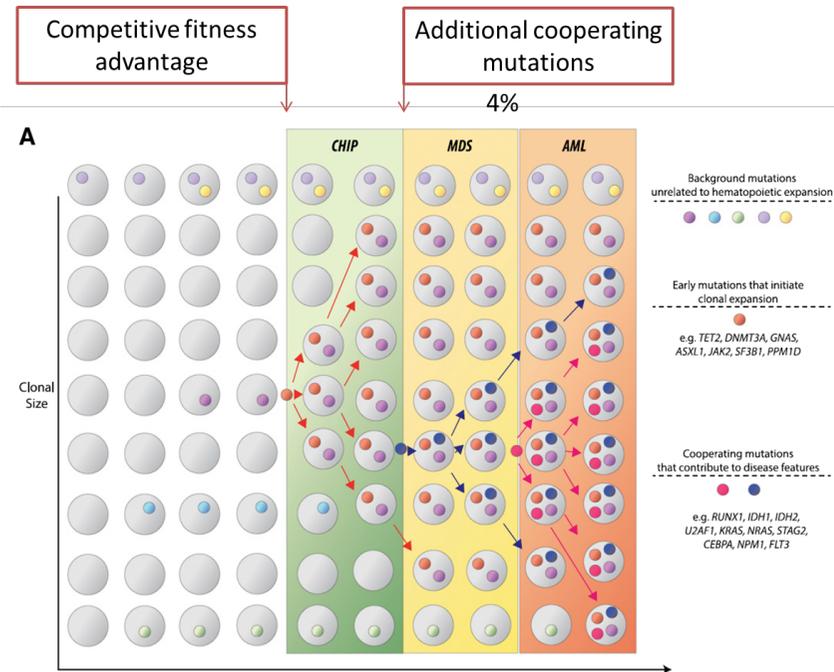
## Variant allele frequency (VAF)



	VAF (%)	APL type	PML-RARA type
<i>RUNX1</i> p.Arg306Cys (rs202037007)	57.57	P	BCR3
<i>TET2</i> p.Val1718Leu (rs142312318)	48.91	S	BCR1
<i>TET2</i> p.Gln810Arg (rs28555446)	50.38	P	BCR3

# Results

## Early mutations vs. germ line mutations



**Table 17. Classification of myeloid neoplasms with germ line predisposition**

**Myeloid neoplasm classification**

**Myeloid neoplasms with germ line predisposition without a preexisting disorder or organ dysfunction**

AML with germ line *CEBPA* mutation

Myeloid neoplasms with germ line *DDX41* mutation\*

**Myeloid neoplasms with germ line predisposition and preexisting platelet disorders**

Myeloid neoplasms with germ line *RUNX1* mutation\*

Myeloid neoplasms with germ line *ANKRD26* mutation\*

Myeloid neoplasms with germ line *ETV6* mutation\*

**Myeloid neoplasms with germ line predisposition and other organ dysfunction**

Myeloid neoplasms with germ line *GATA2* mutation

Myeloid neoplasms associated with BM failure syndromes

Myeloid neoplasms associated with telomere biology disorders

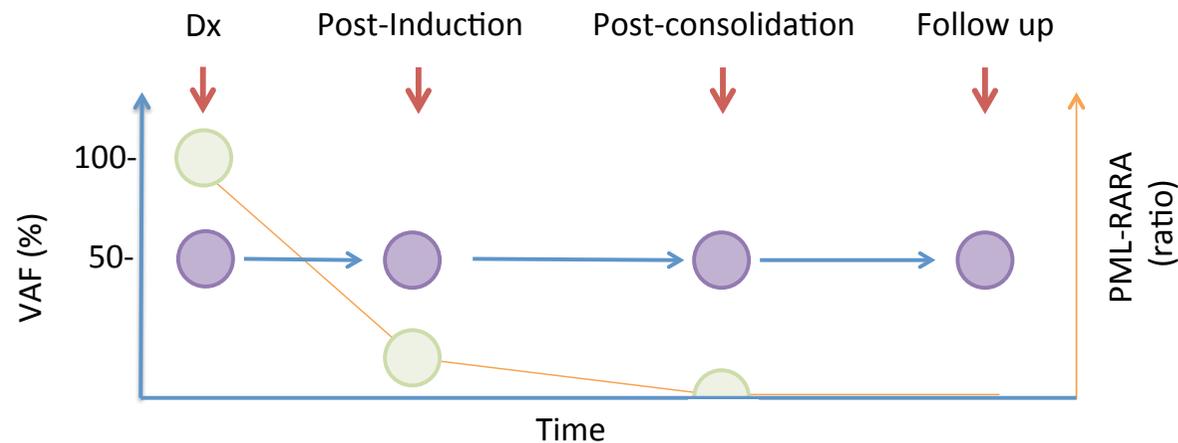
JMML associated with neurofibromatosis, Noonan syndrome or

Noonan syndrome-like disorders

Myeloid neoplasms associated with Down syndrome\*

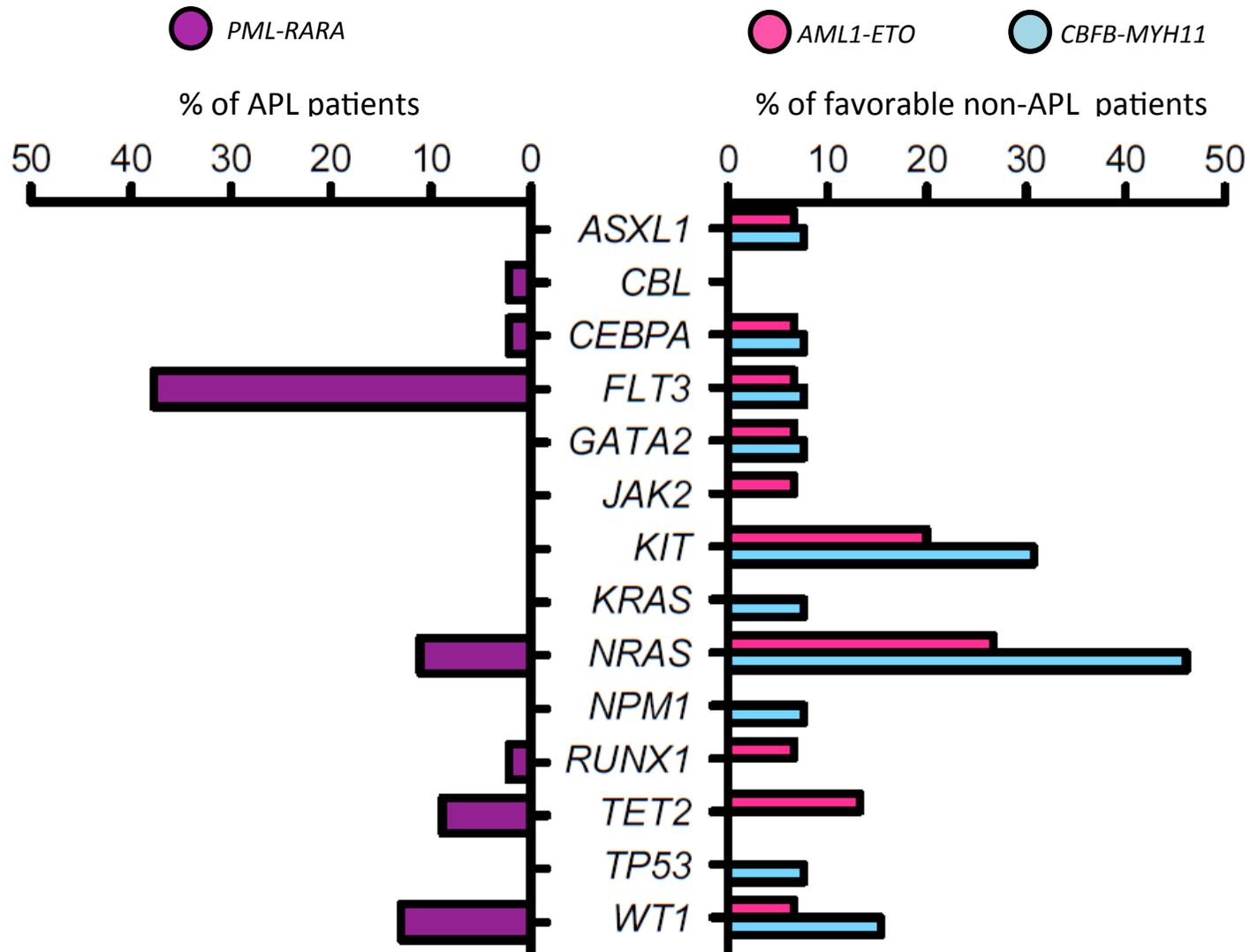
\*Lymphoid neoplasms also reported.

Arber et al 2016



# Results

## Mutational patterns in APL vs. other favourable risk AML



# Conclusions

- Our data shows that NGS is a valid method to detect recurrently mutated genes in AML.
- APL patients harbor somatic mutations in *FLT3*, *WT1*, and *NRAS*.
- Mutations in genes involved in signaling processes are usually found at a low VAF, and patients harboring them could benefit of combined targeted therapy.
- APL patients harbor germ-line mutation in *TET2* and *RUNX1*.
- The mutational spectrum of APL is different from that of other favourable AML
- Further research is needed in order to understand the involvement of these mutations in the clinical management of APL.

Thank you for your attention!



The Research Group in Haematology and Haemotherapy

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