

Come potenziare la terapia con immune checkpoint inhibitors

Massimo Massaia

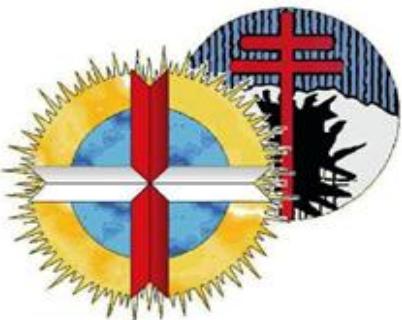
SC Ematologia - AO S. Croce e Carle – Cuneo, Italy

Laboratorio di Immunologia dei Tumori del Sangue, CeRMS - Torino, Italy

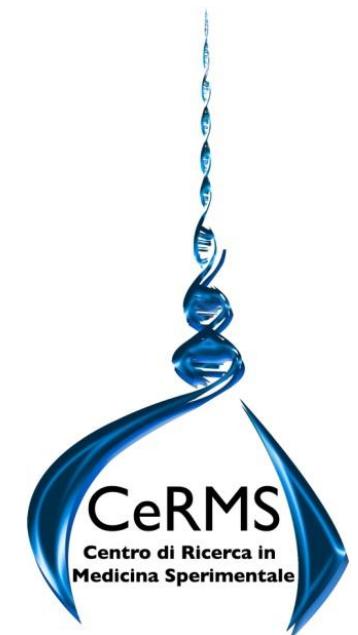
HIGHLIGHTS IN EMATOLOGIA

Treviso, Ospedale Ca' Foncello

22-23 Novembre 2019



AO S.Croce e Carle
Cuneo



DISCLOSURES:

MASSIMO MASSAIA

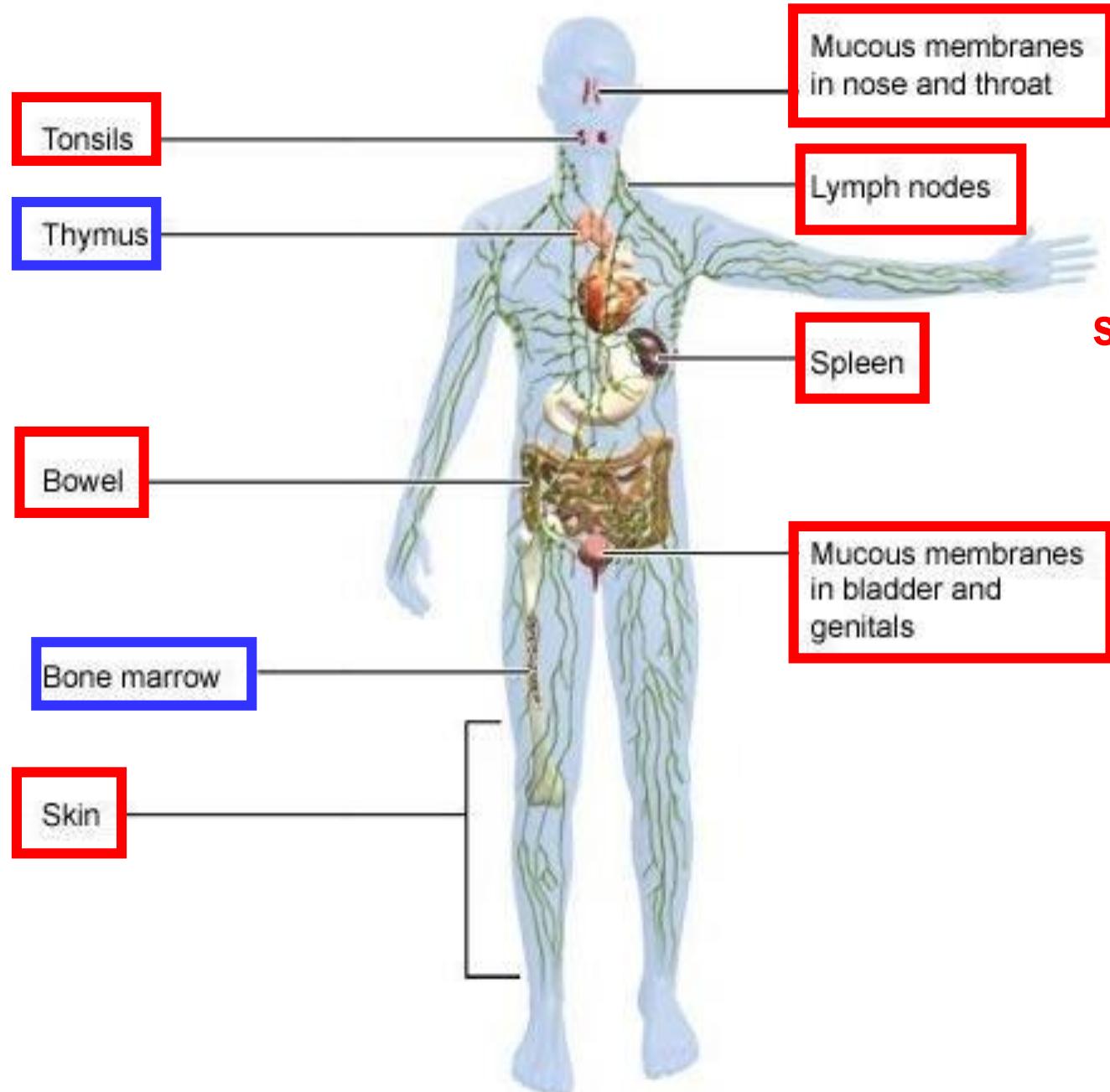
Come da nuova regolamentazione della Commissione Nazionale per la Formazione Continua del Ministero della Salute, è richiesta la trasparenza delle fonti di finanziamento e dei rapporti con soggetti portatori di interessi commerciali in campo sanitario.

- Fondi per la ricerca da aziende con interessi commerciali in campo sanitario: *Gilead, Novartis*
- Partecipazione ad Advisory Board: *Abbvie, Janssen*

The human immune system

primary lymphoid organs

secondary lymphoid organs



The main functions of the immune system

Immune system belongs to the basic homeostatic mechanisms

Defense - identification and protection against pathogenic microorganisms and their toxins

Autotolerance – recognition of own tissues and keeping tolerance to them

Immune surveillance - identifying and removing the old , damaged and otherwise changed cells

Internal Threat

Autoimmune problem

(*type 1 diabetes, rheumatoid arthritis, psoriasis, multiple sclerosis, lupus, inflammatory bowel disease*)

External Threat

Allergic Reaction

(*hay fever, eczema, asthma, sinusitis*)

Immune Over-reaction

BALANCED IMMUNE SYSTEM = OPTIMAL EFFECTIVENESS

Immune Under-reaction

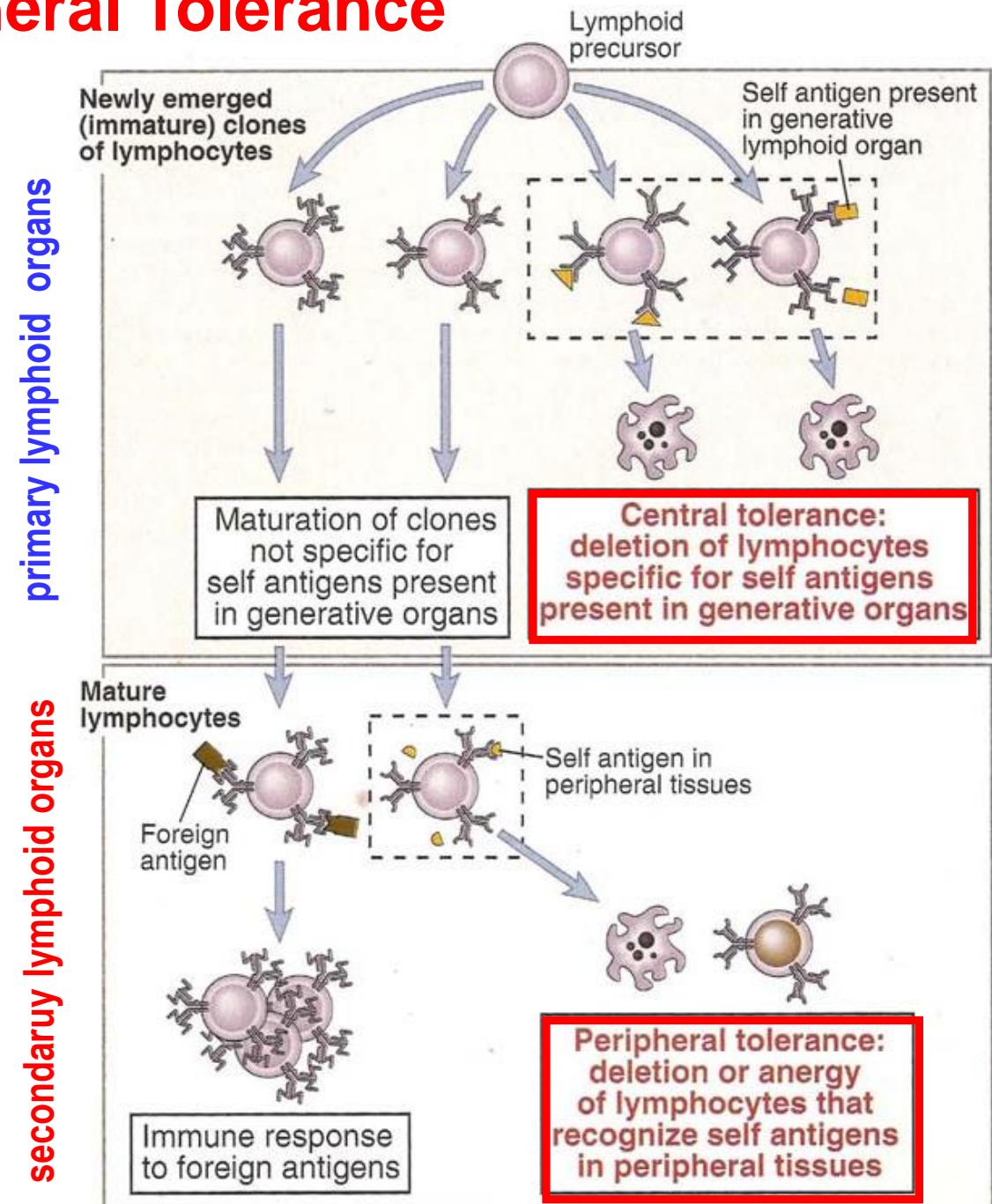
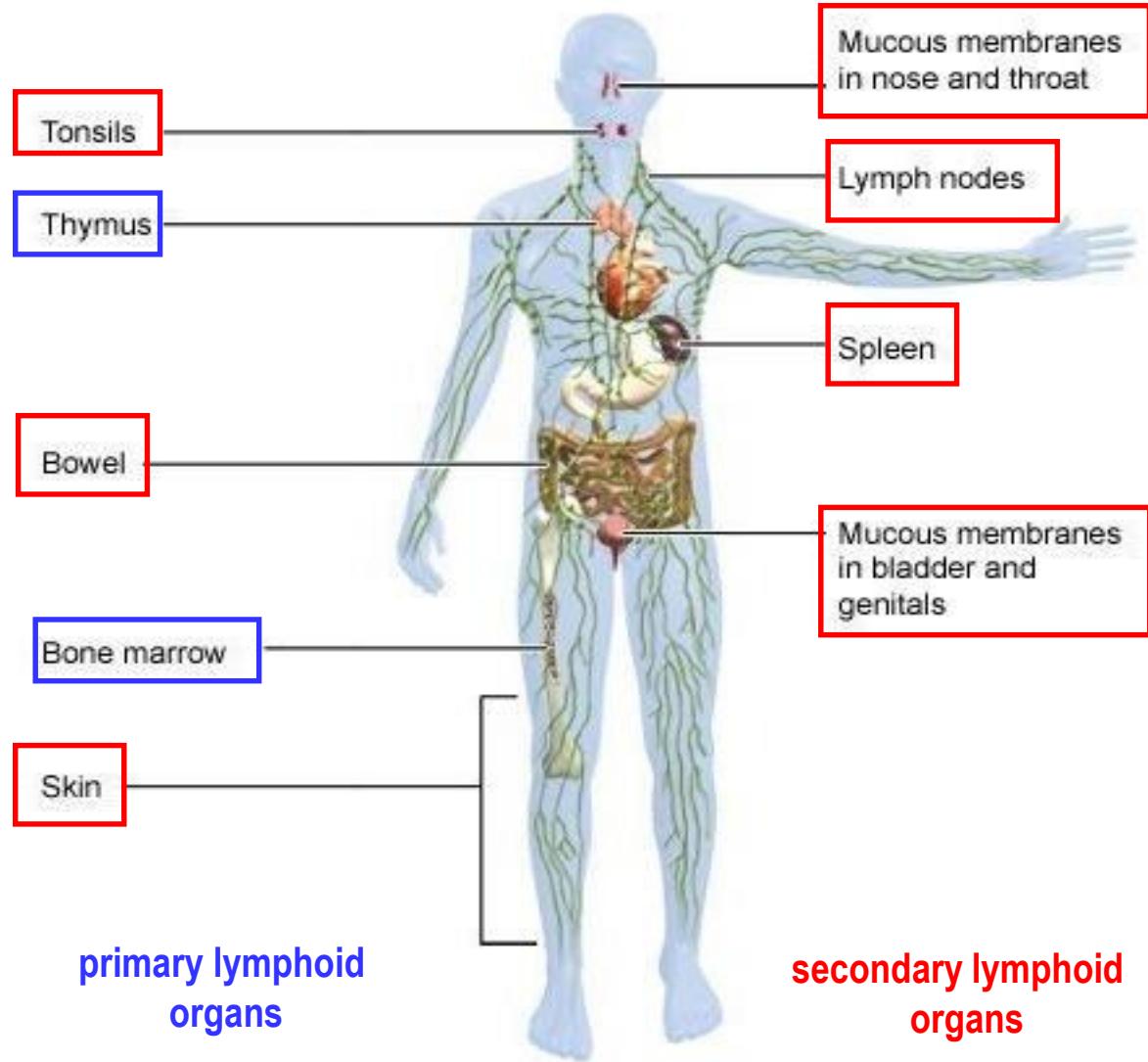
Cancer

(*hepatitis, HIV, shingles, TB*)

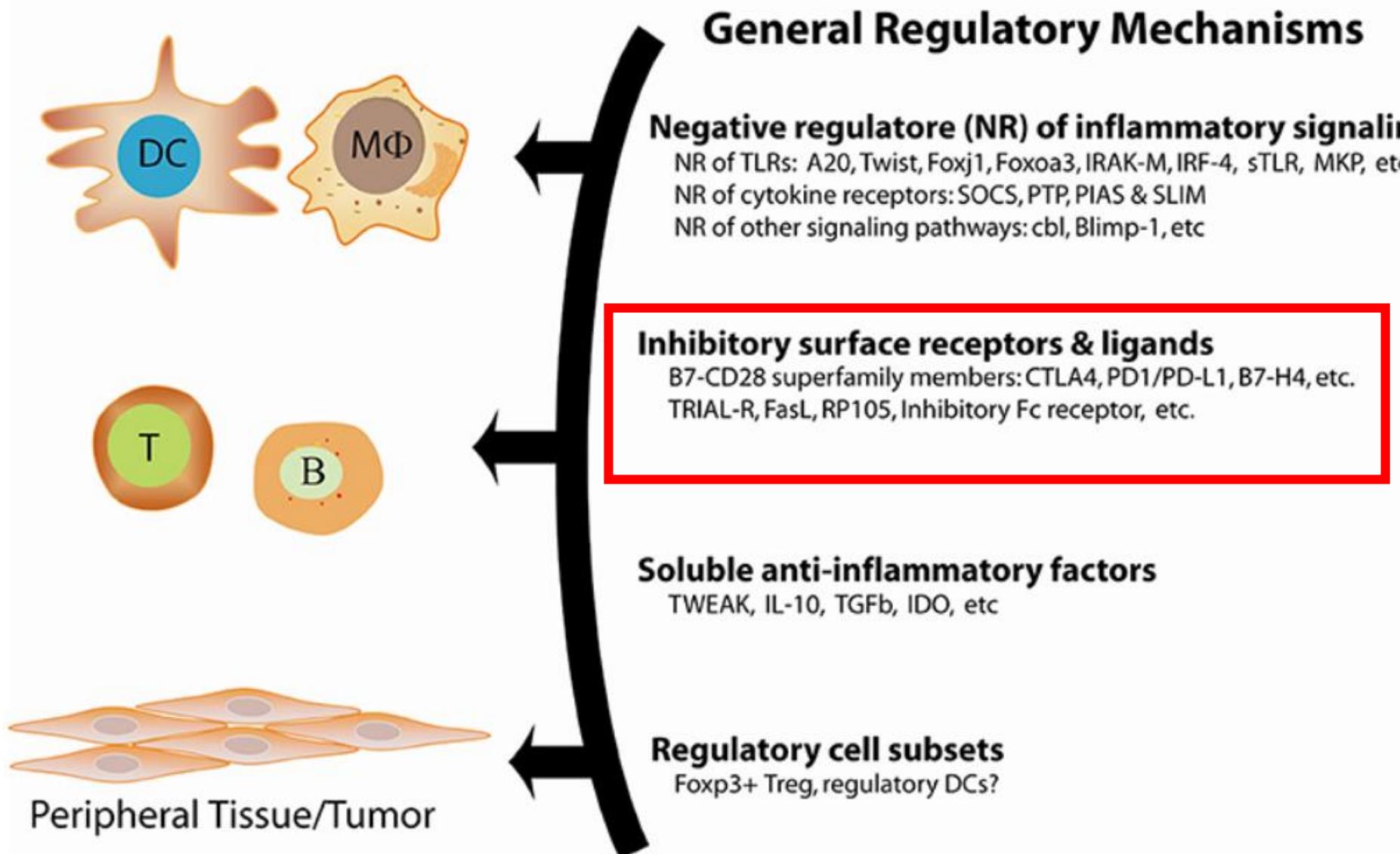
Infection

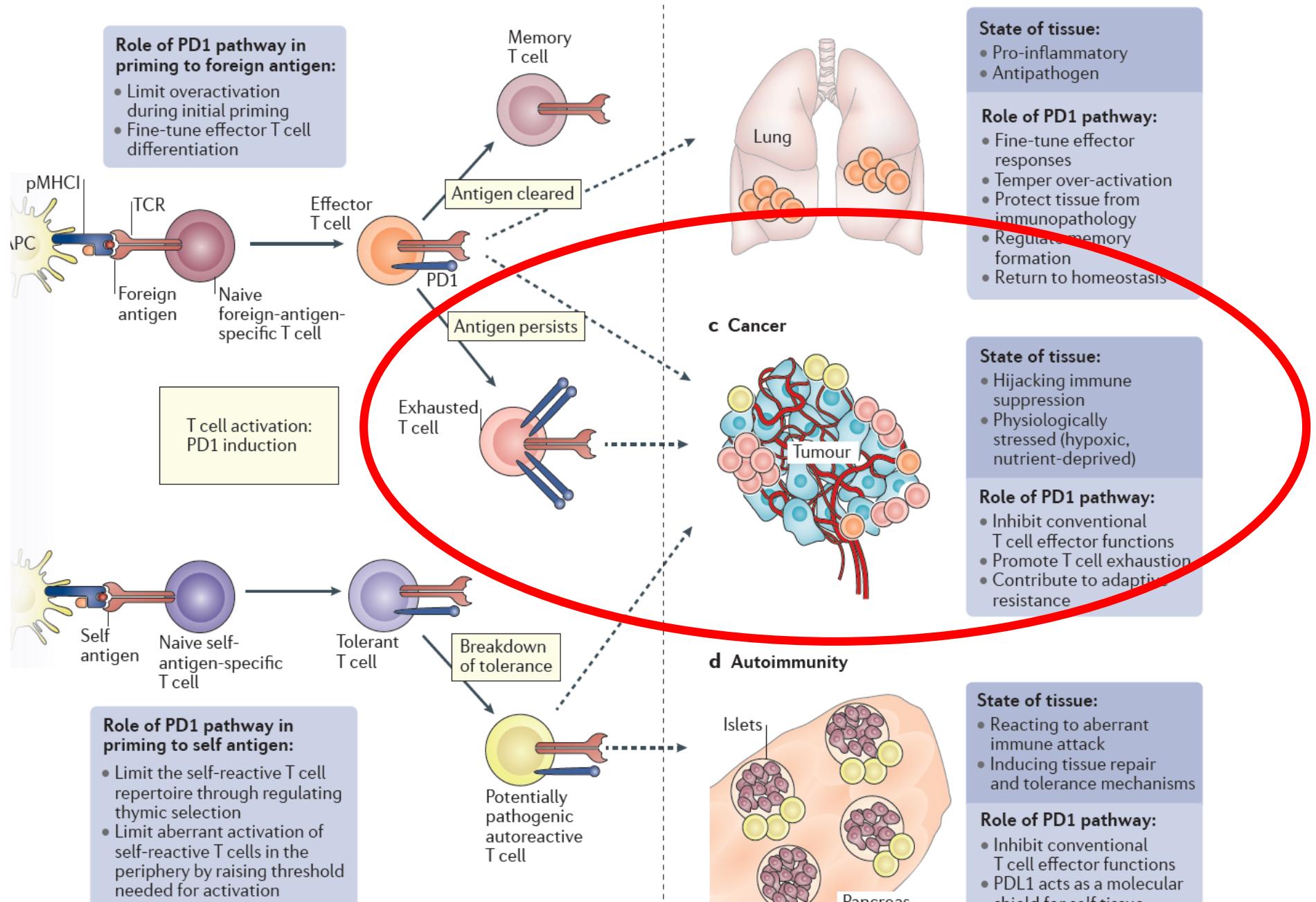
(*viruses, bacteria, fungi and parasites*)

Central and Peripheral Tolerance



Peripheral tolerance maintained by multiple negative regulatory mechanisms controlling multiple levels and phases of immune responses





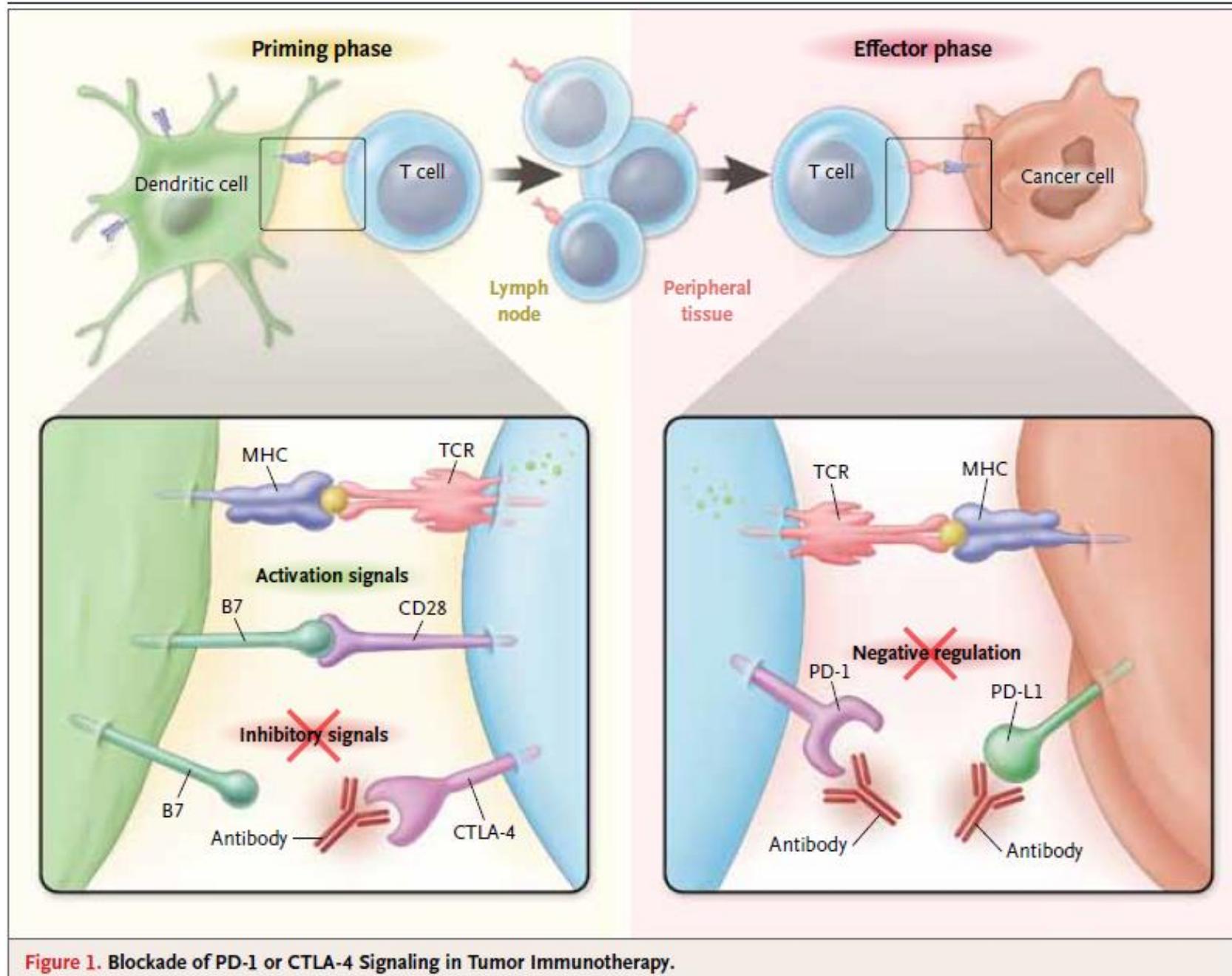
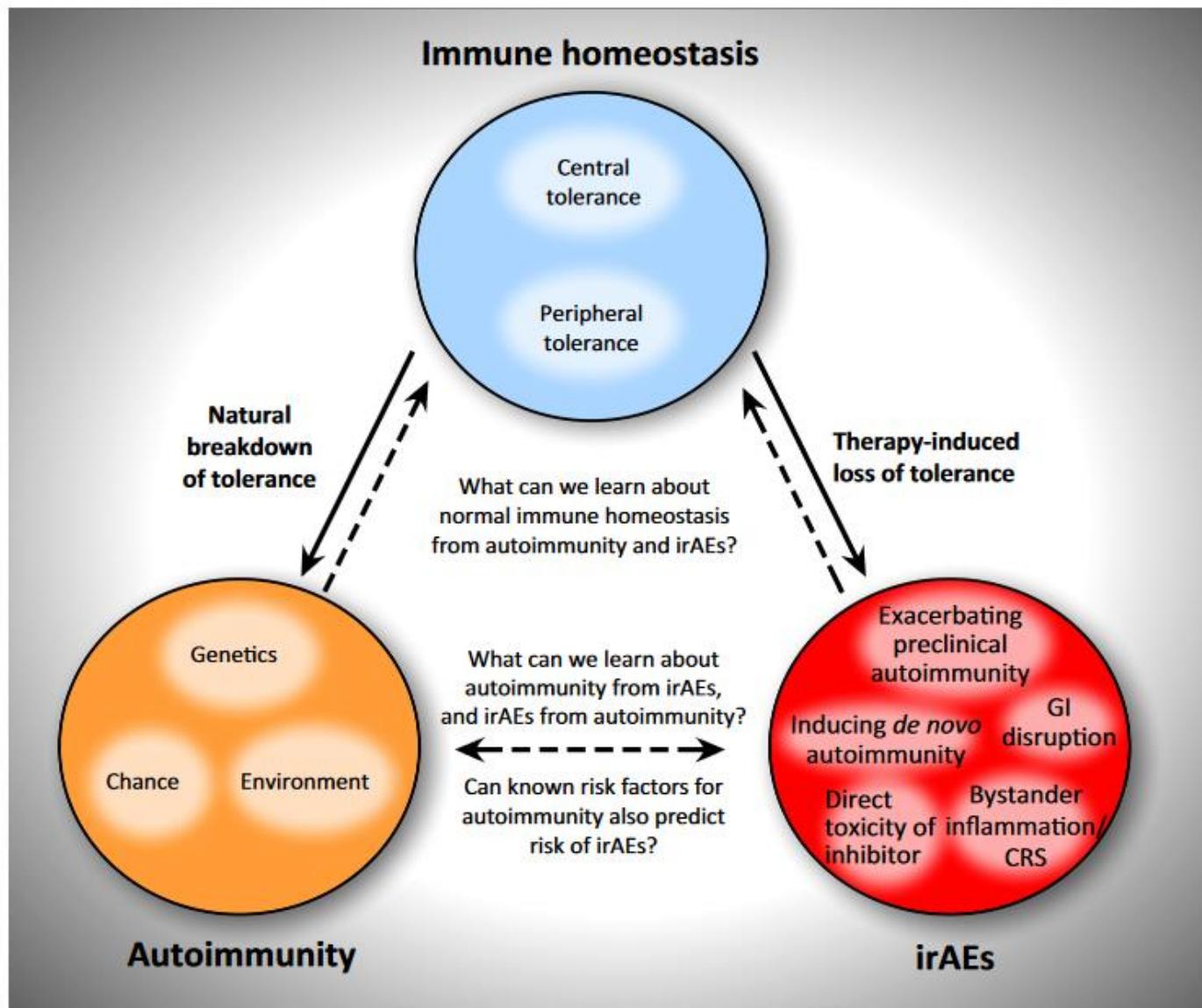


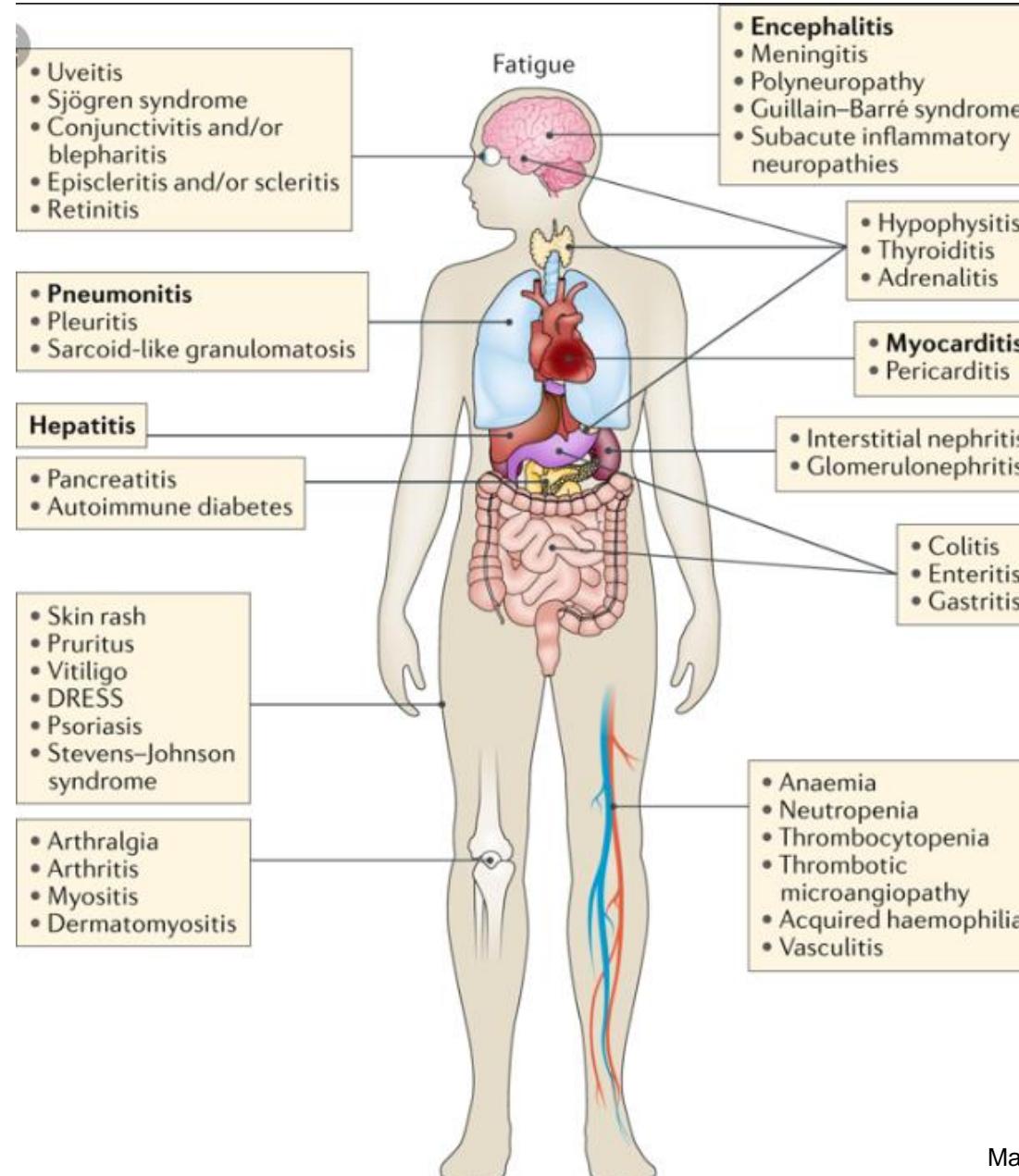
Figure 1. Blockade of PD-1 or CTLA-4 Signaling in Tumor Immunotherapy.

Key Figure

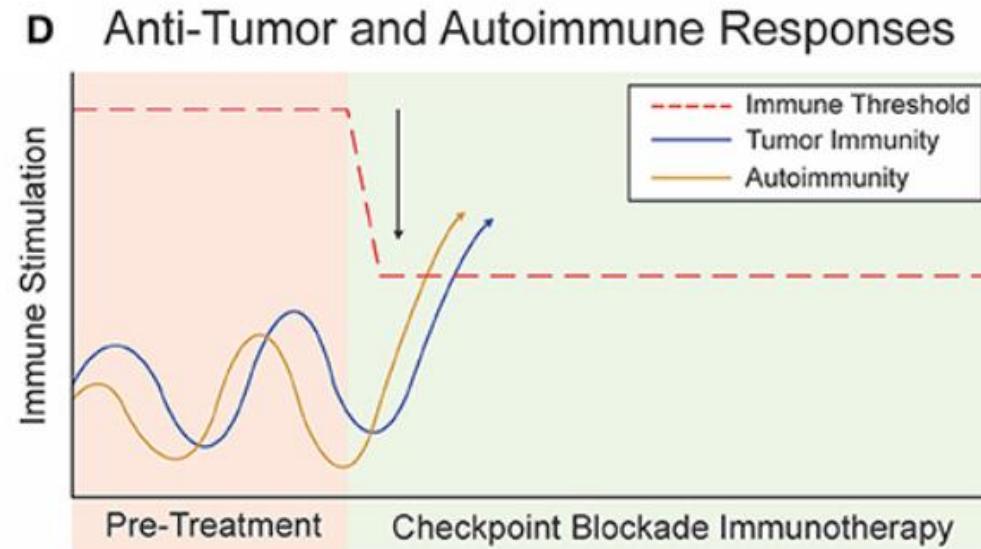
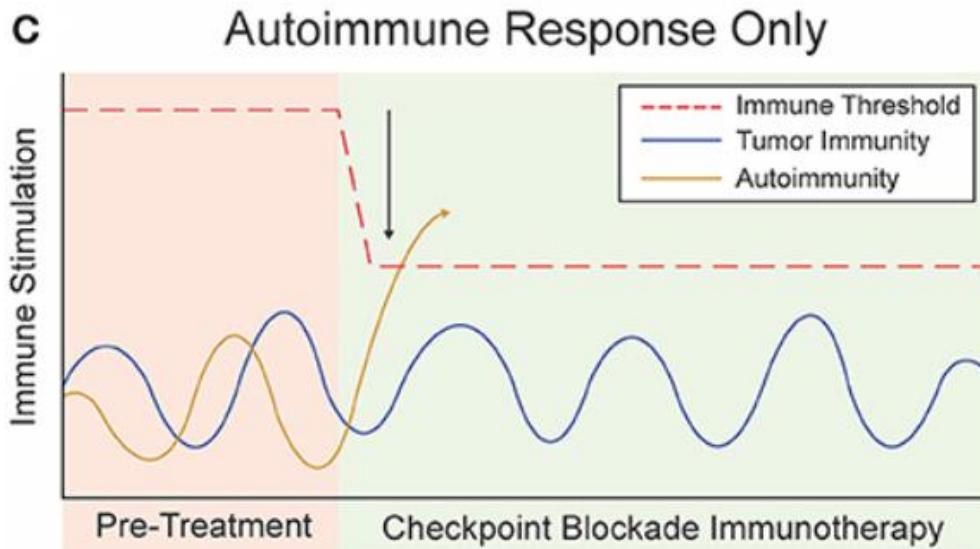
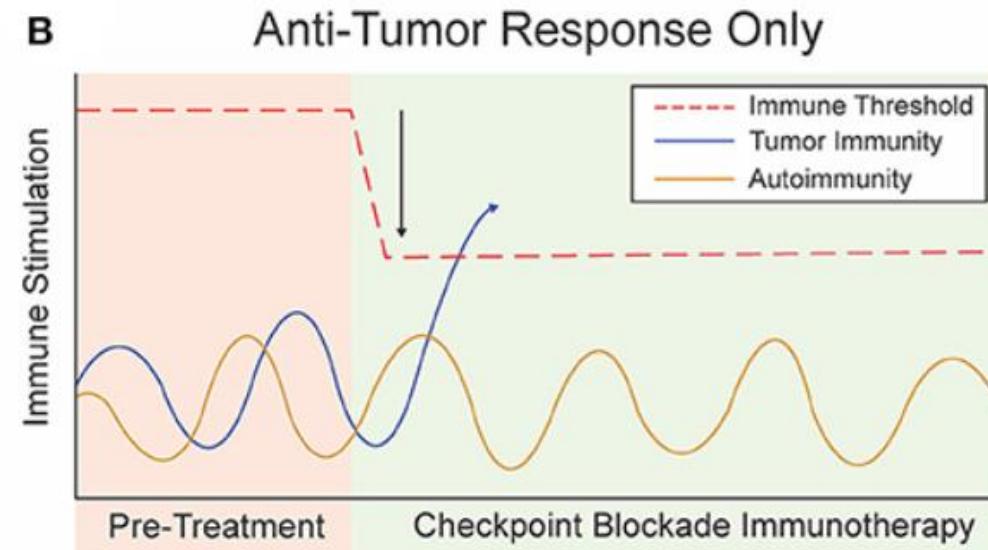
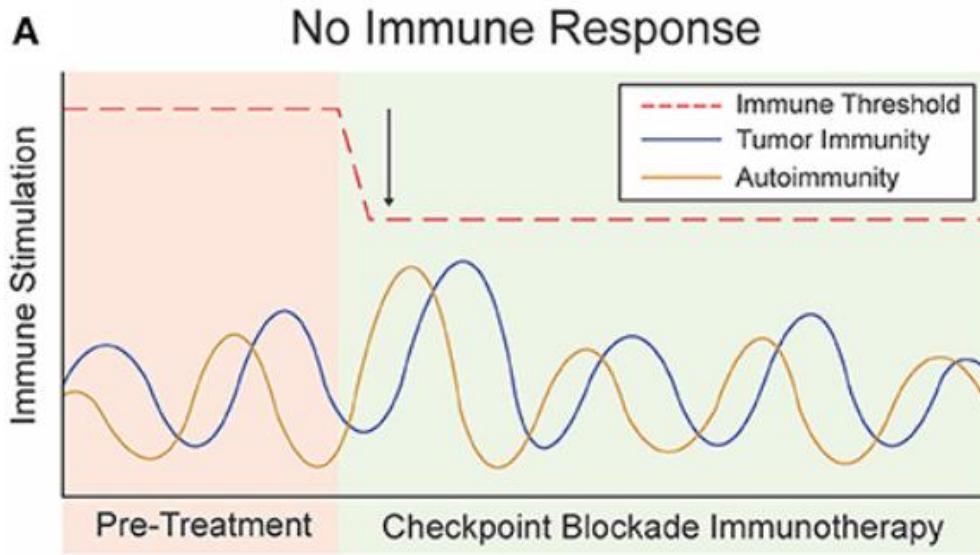
Pathogenic Immune Responses and Onset of Autoimmunity or irAEs



The spectrum of IRAEs induced by ICP blockade



A Threshold Model for Immune Activation



REVIEW

Open Access

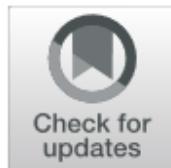
Immune-related adverse events and anti-tumor efficacy of immune checkpoint inhibitors



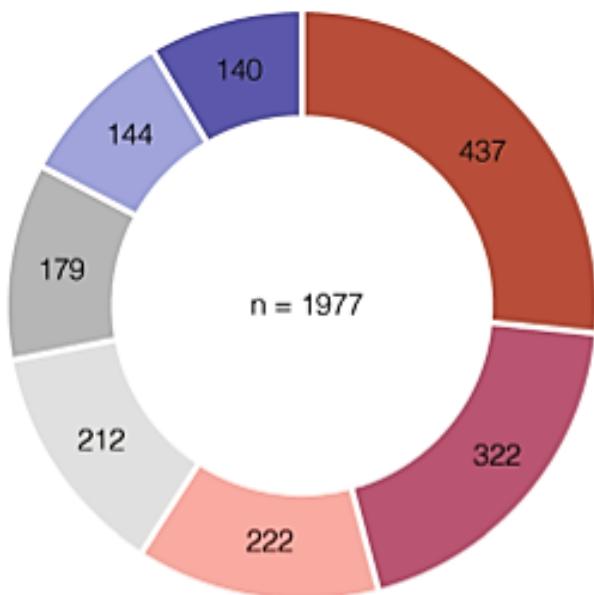
Satya Das* and Douglas B. Johnson

ORR, PFS and OS mucch better in IRAES+ vs IRAES- pts after treatment with α-PD-1/PD-1L

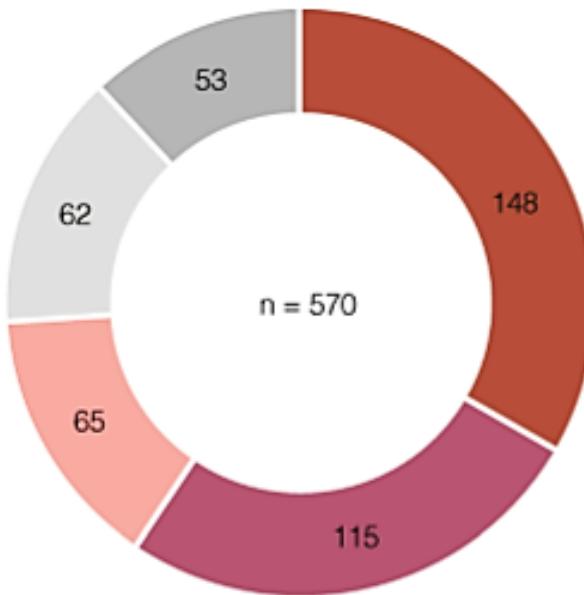
Advances in cancer immunotherapy 2019 – latest trends

**A**

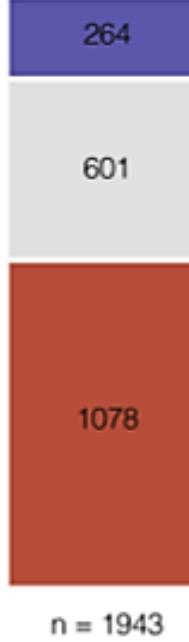
Anti-PD-1-PD-L1 trials by cancer type

**B**

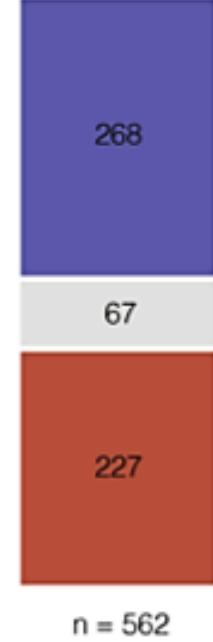
CAR T cell trials by cancer type

**C**

Anti-PD-1-PD-L1 trials by region



CAR T cell trials by region



■ Lung cancers ■ Urological cancers
■ GI cancers ■ Breast cancers
■ Melanoma ■ Lymphomas
■ HN cancers

■ B Cell lymphoma ■ Multiple Myeloma
■ ALL ■ GI cancers
■ CLL

■ USA
■ Europe
■ China



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Programmed cell death protein receptor and ligands in haematological malignancies – Current status



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^c Dept. of Cancer Prevention and Therapy, Wrocław Medical University, Poland

Table 1

Clinical trials with use of PD-1 or PD-L1 inhibitors in haematological malignancies.

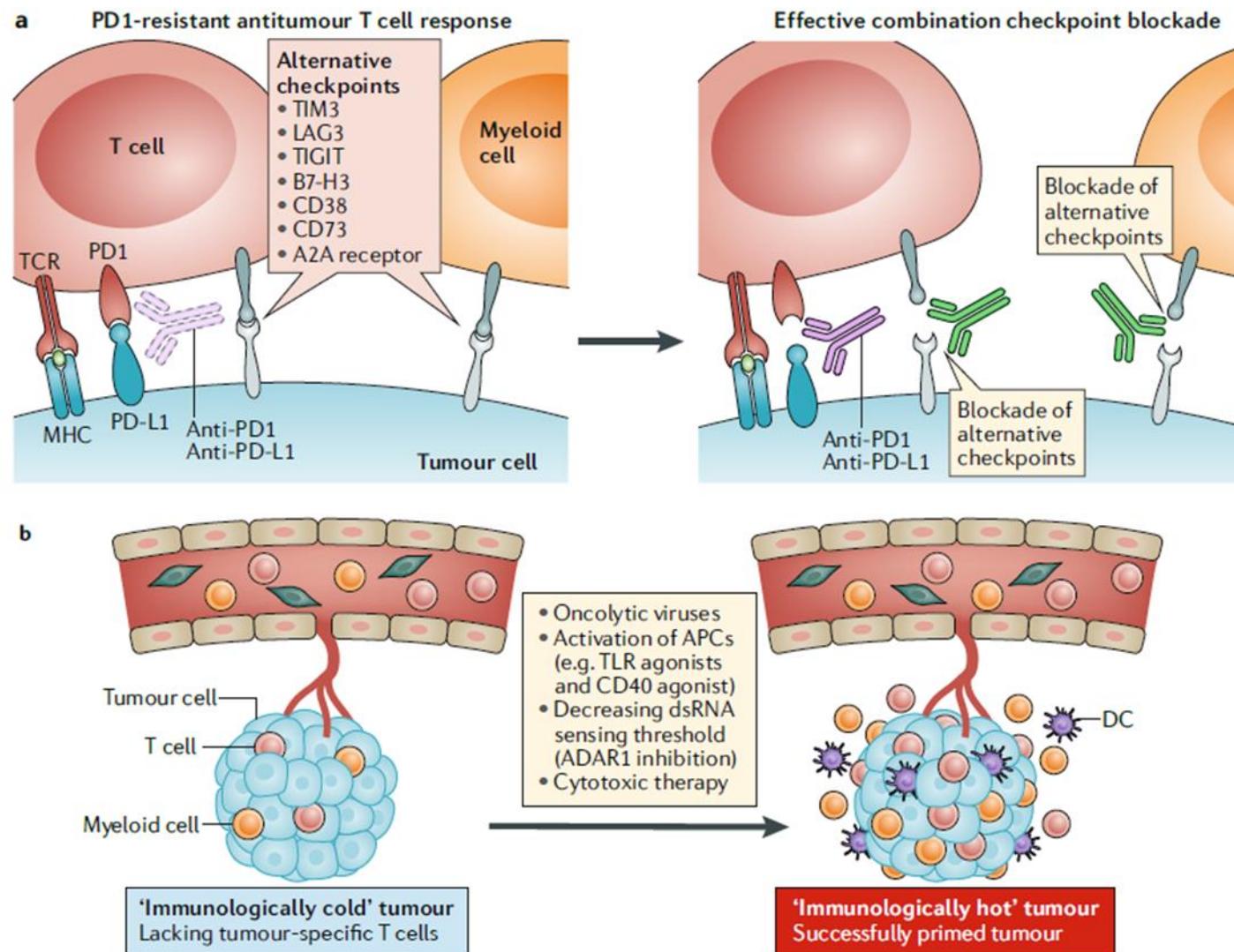
Disease	PD-1 or PD-L1 inhibitor	Phase of clinical trial	Author	Clinical outcome
Multiple myeloma	Pembrolizumab + Lenalidomide + Low-Dose Dexamethasone for Relapsed/Refractory Multiple Myeloma	I	Mateos et al.	ORR = 76% VGPR = 24% PR = 53% CR = 94%
	Pembrolizumab + Pomalidomide + Low-Dose dexamethasone for relapsed/refractory multiple myeloma	II	Bardos A. et al.	ORR = 60% sCR = 6% CR = 21% OS = 174 months
Chronic Lymphocytic Leukemia	Pembrolizumab in chronic lymphocytic leukemia with Richter's transformation (RT) and relapsed CLL	II	Ding W. et al.	ORR in patients with RT = 44% ORR in CLL patients = 0% OS in patients with RT = 107 months
Relapsed or Refractory Hematologic Malignancy:	Nivolumab	Ib	Lesokhin, A.M et al.	ORR for FL = 40% ORR for DLBCL = 36% ORR for other BCL = 15% ORR for MF = 40%
	Nivolumab + Ipilimumab	Ib	Ansell S. et al.	ORR for HL = 74% ORR for B-cell NHL = 20% ORR for T-cell NHL = 9% ORR for MM = 0%
	Pidilizumab	I	Berger R. et al.	ORR = 33%
Acute myeloid leukemia (AML)	Nivolumab + Azacitidine in with Relapsed Acute Myeloid Leukemia (AML)	II	Dauer N. et al.	ORR = 69% CR = 18% Median OS = 9,3 months
Myelodysplastic syndrome (MDS)	Nivolumab with Azacitidine in untreated patients with Myelodysplastic Syndromes and In comparison vs Nivolumab as single after treatment wih Hypomethylating Agents	II	Garcia-Manero G. et al.	ORR in AZA + Nivo = 69% CR in AZA + Nivo = 15% HI in AZA + Nivo = 15% Nivo vs Ipi monotherapy ORR = 0% vs 22% (p = 0.156)
	Pembrolizumab	II	Garcia-Manero G. et al.	ORR = 4% PR = 4% Marrow CR = 11% HI = 11% Overall Survival Rate after 24 weeks = 49%
Hodgkin lymphoma (HL)	Nivolumab in Relapsed or Refractory Hodgkin's lymphoma	I	Ansell S. et al.	ORR = 87% CR = 17% PFS at 24 weeks = 86%
	Nivolumab in Relapsed or Refractory Hodgkin's Lymphoma	II	Timmerman J.M et al.	ORR = 68% CR = 8% PFS after 6 months = 77% PFS after 12 months = 54%
	Nivolumab in Japanese patients with relapsed or refractory classical Hodgkin lymphoma.	II	Maruyama D. et al.	ORR = 81.3% CR = 23% PR = 53% OS after 6 months = 100%
	Pembrolizumab in Patients With Classical Hodgkin Lymphoma After Brentuximab Vedotin Failure	I	Armand P. et al.	ORR = 65% CR = 16% PFS rate after 24 weeks 69% PFS rate after 52 weeks = 46%
Diffuse large B-cell lymphoma (DLBCL)	Pembrolizumab for Relapsed/Refractory Classic Hodgkin Lymphoma.	II	Chen R. et al.	ORR = 69% CR = 224%
	Pidilizumab after autologous hematopoietic stem-cell transplantation for diffuse large B-cell lymphoma	II	Armand P. et al.	ORR = 51% PFS rate after 16 months = 70% CR = 34%
	Pembrolizumab in relapsed/refractory primary mediastinal large B-cell lymphoma.	Ib	Zinzani PL. Et al.	ORR = 41% CR = 12%
Non-Hodgkin lymphoma (FL + DLBCL)	Atezolizumab + Obinutuzumab in relapsed /refractory NHL	Ib	Palomba M.L. et al.	ORR for FL = 57% ORR for DLBCL = 16%
Follicular lymphoma	Pidilizumab + Rituximab in relapsed follicular lymphoma	II	Westin J.R. et al.	ORR = 66% CR = 52%
Mycosis Fungoïdes and Sezary Syndrome	Pembrolizumab in Relapsed/Refractory Mycosis Fungoïdes and Sezary Syndrome	II	Khodadoust M. et al.	ORR = 38% CR = 4% PFS rate after one year = 69%

Hodgkin lymphoma - PMBCL - Richter syndrome - PTCL/AL

ICP blockade in myeloid malignancies: a promise under investigation

Intervention	Outcomes
Nivolumab + 5-AZA	75% CR/CRp; 50% 1-year survival
Nivolumab + 5-AZA	ORR: 33% (22% CR/CRi); median OS 6.3 months
Idarubicin + cytarabine ± nivolumab	77% CR/CRi; median OS 18.54 (nivolumab group) vs 13.2 months (I + A alone), p = 0.2
Pembrolizumab + decitabine	1 MRD-negative CR; median OS 7 months
Ipilimumab + 5-AZA	71% CR/CRp; 68% 1-year survival

Mechanisms of innate and acquired resistance to immune checkpoint inhibition (I)



Mechanisms of innate and acquired resistance to immune checkpoint inhibition (III)

a Oncogenic signalling pathways

MAPK signalling

- Increased production of immuno-suppressive cytokines IL-6 and IL-10
- Negative regulation of antigen presentation
- Suppression of differentiation antigens (melanoma)
- Reduced sensitivity to antiproliferative effects of IFN γ and TNF

WNT- β -catenin signalling

- Increased production of immunosuppressive cytokines
- Disruption of BATF3 $^+$ dendritic cell recruitment by CCL4
- T_{reg} cell development

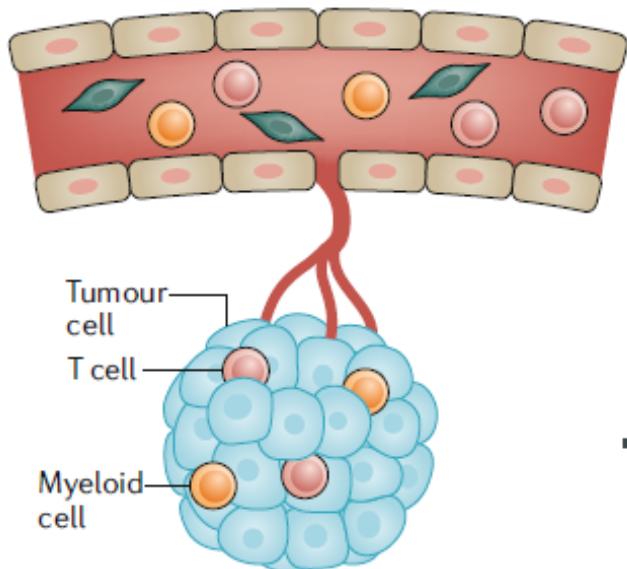
CDK4-CDK6 signalling

- Decreased sensitivity to dsRNA via DNMT1
- Decreased antigen presentation
- Decreased interferon target gene activation

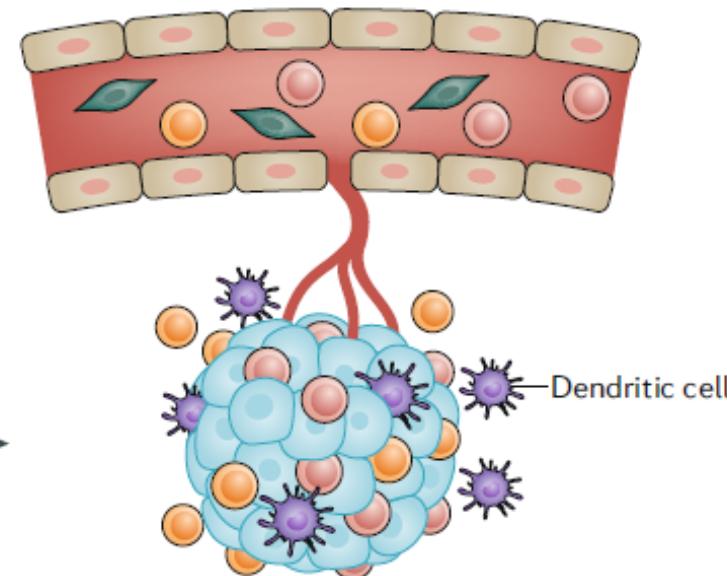
Pathways activated after PTEN loss

- Diminished type I interferon response to PAMPs
- Poor T cell recruitment via activation of autophagosome

b



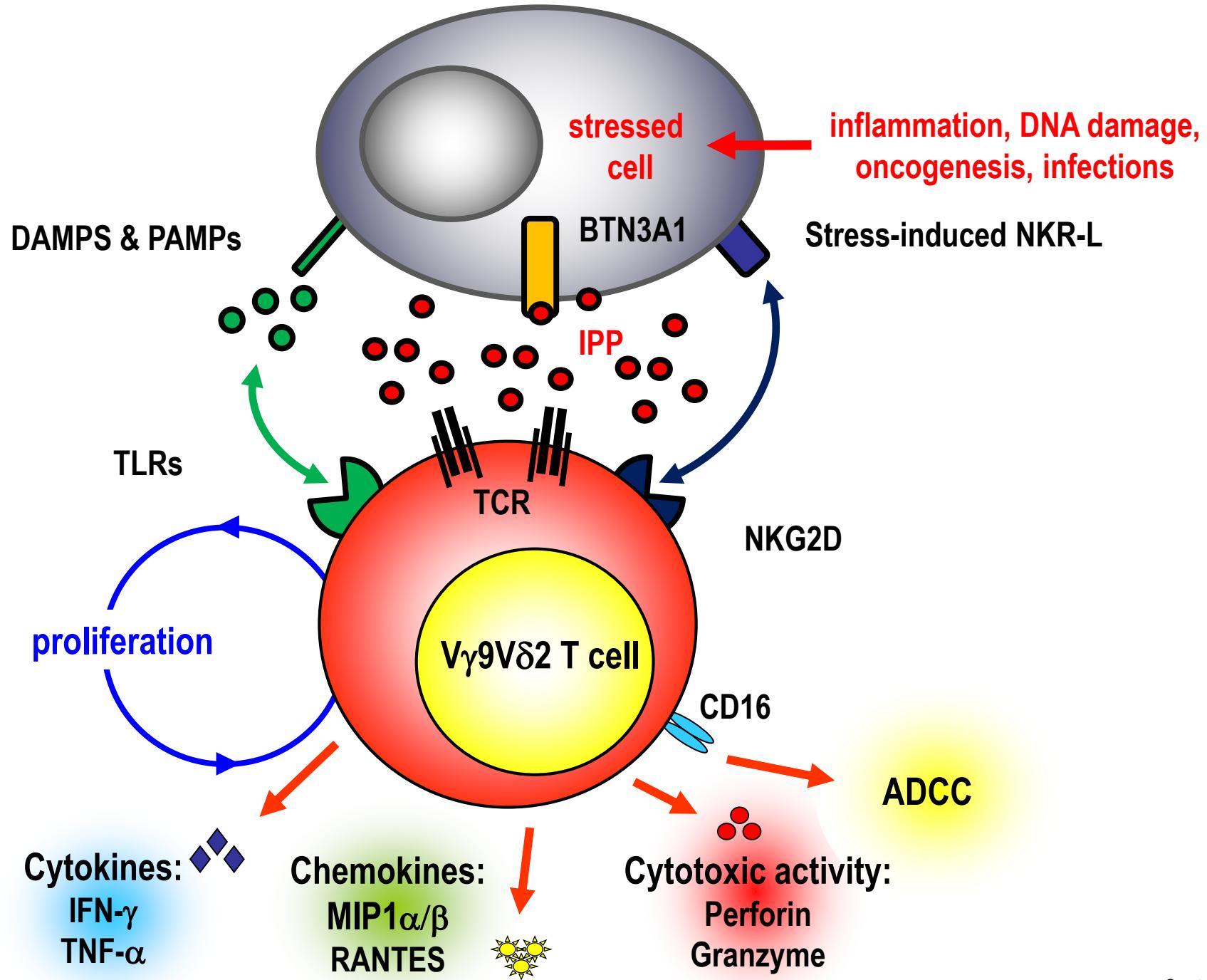
- Inhibition of CDK4-CDK6 signalling
- Inhibition of WNT signalling
- Inhibition of MAPK (BRAF)

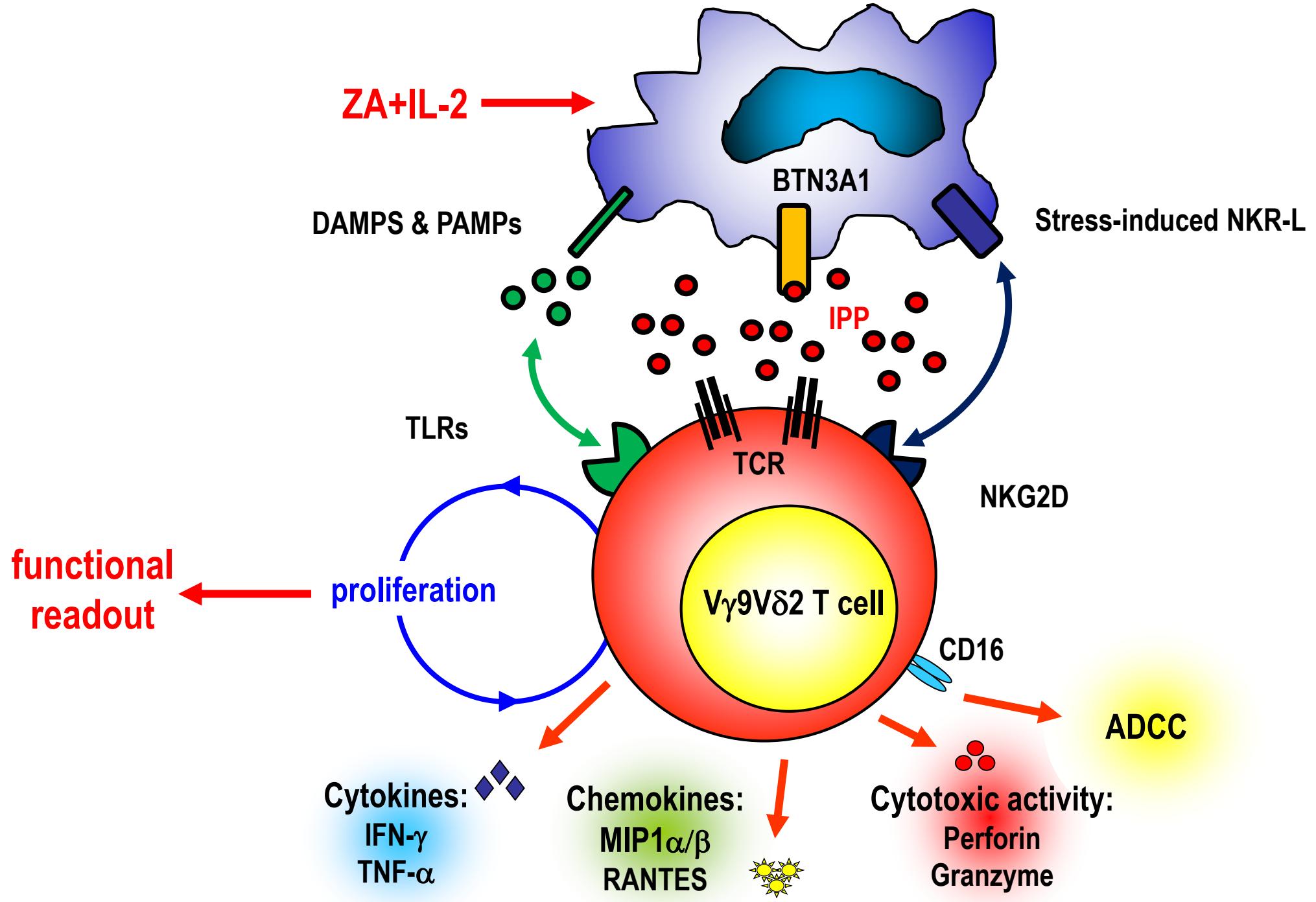


- Dendritic cell activation
- T cell infiltration
- Increased tumour antigen presentation
- Increased dsRNA, IFN γ and TNF sensitivity

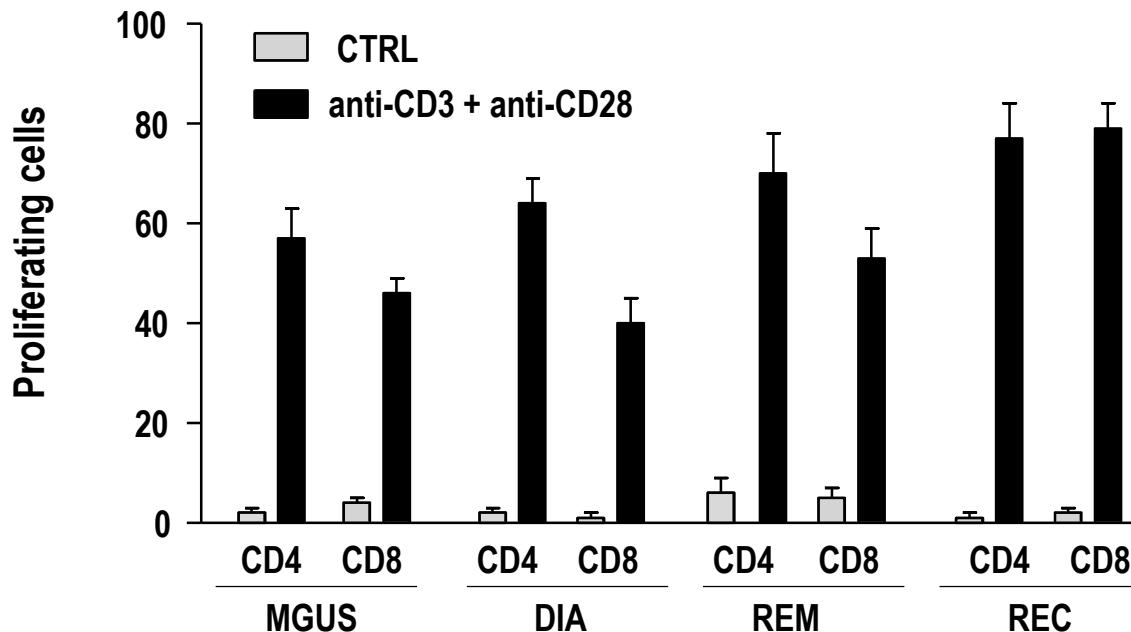
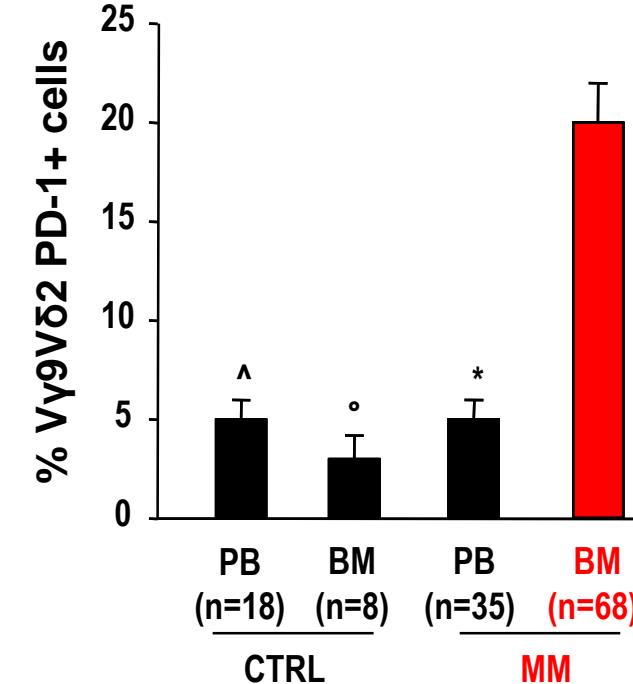
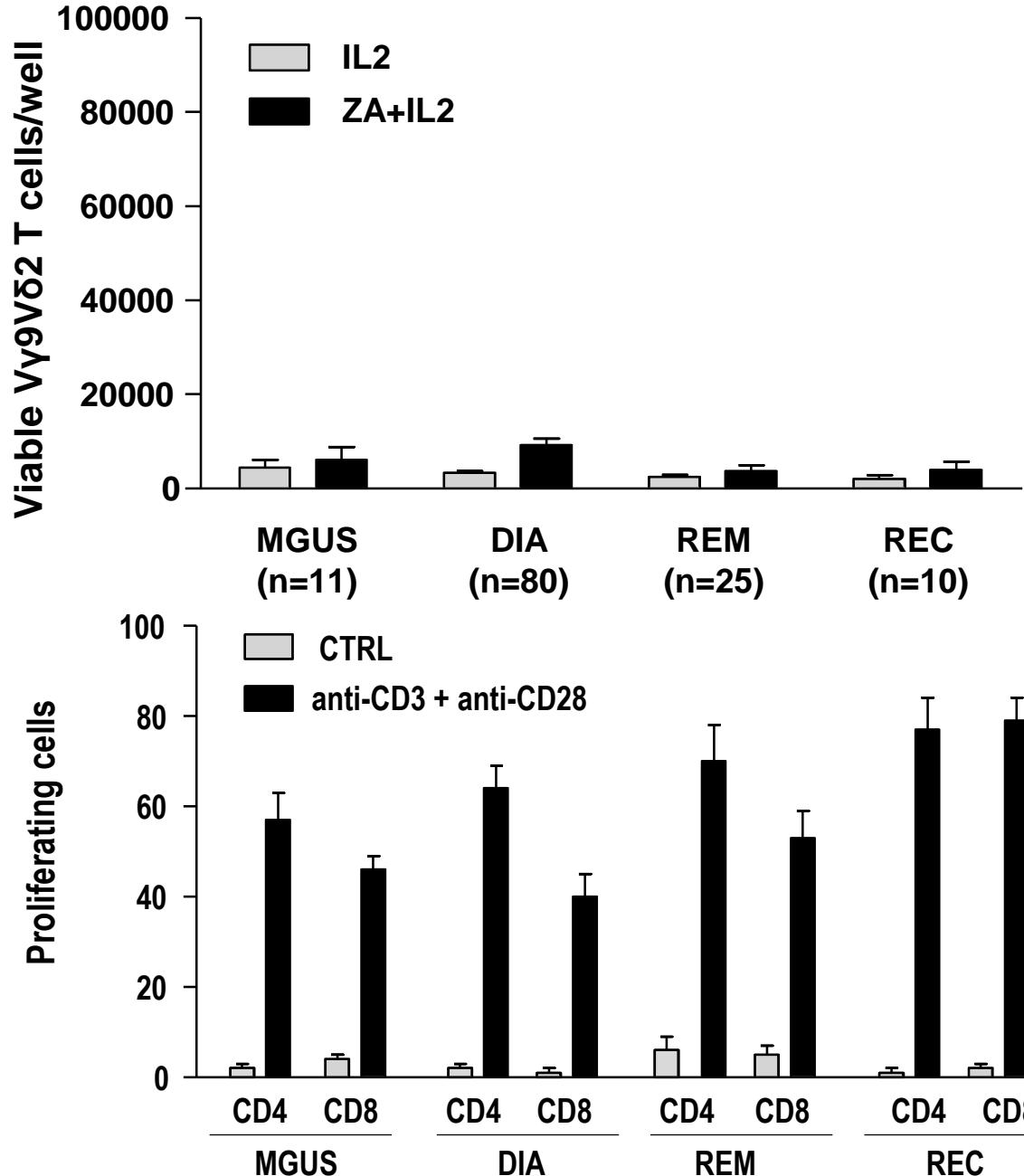
V γ 9V δ 2 T cells as cellular decoders of the immune suppression network in the BM of MM patients



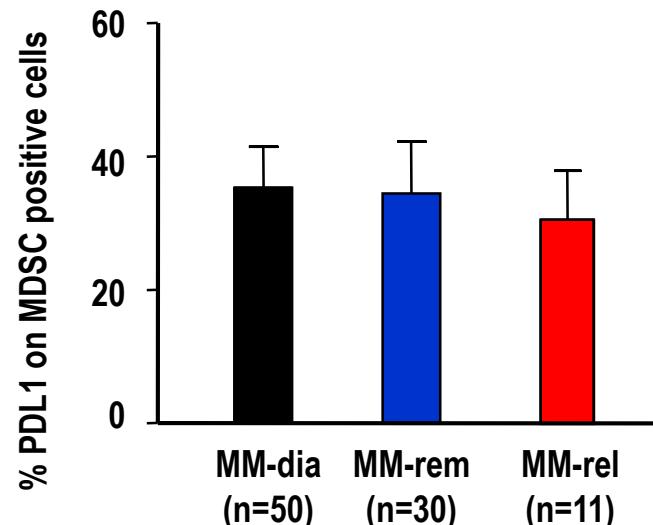
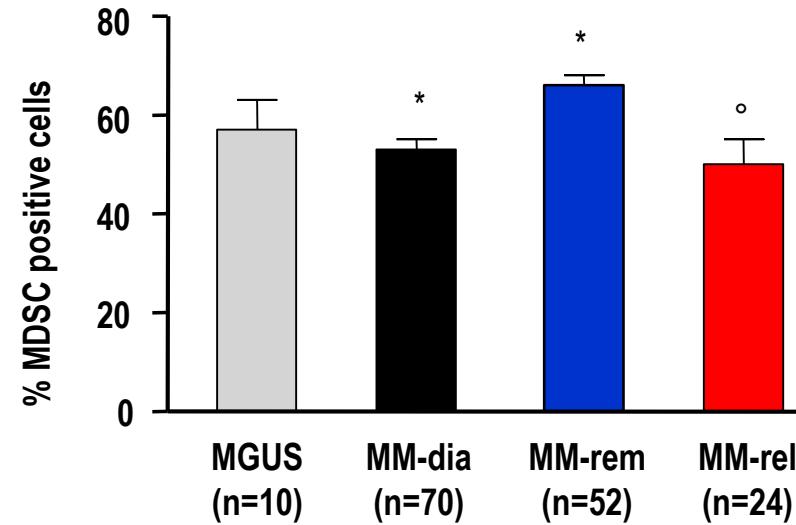
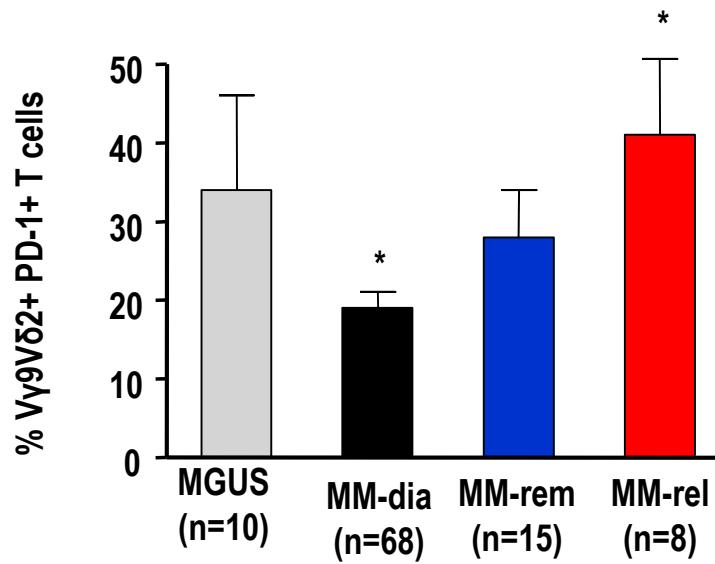




MM BM V γ 9V δ 2 T cells are anergic to pAg stimulation



Early and long-lasting immune suppressive TME commitment

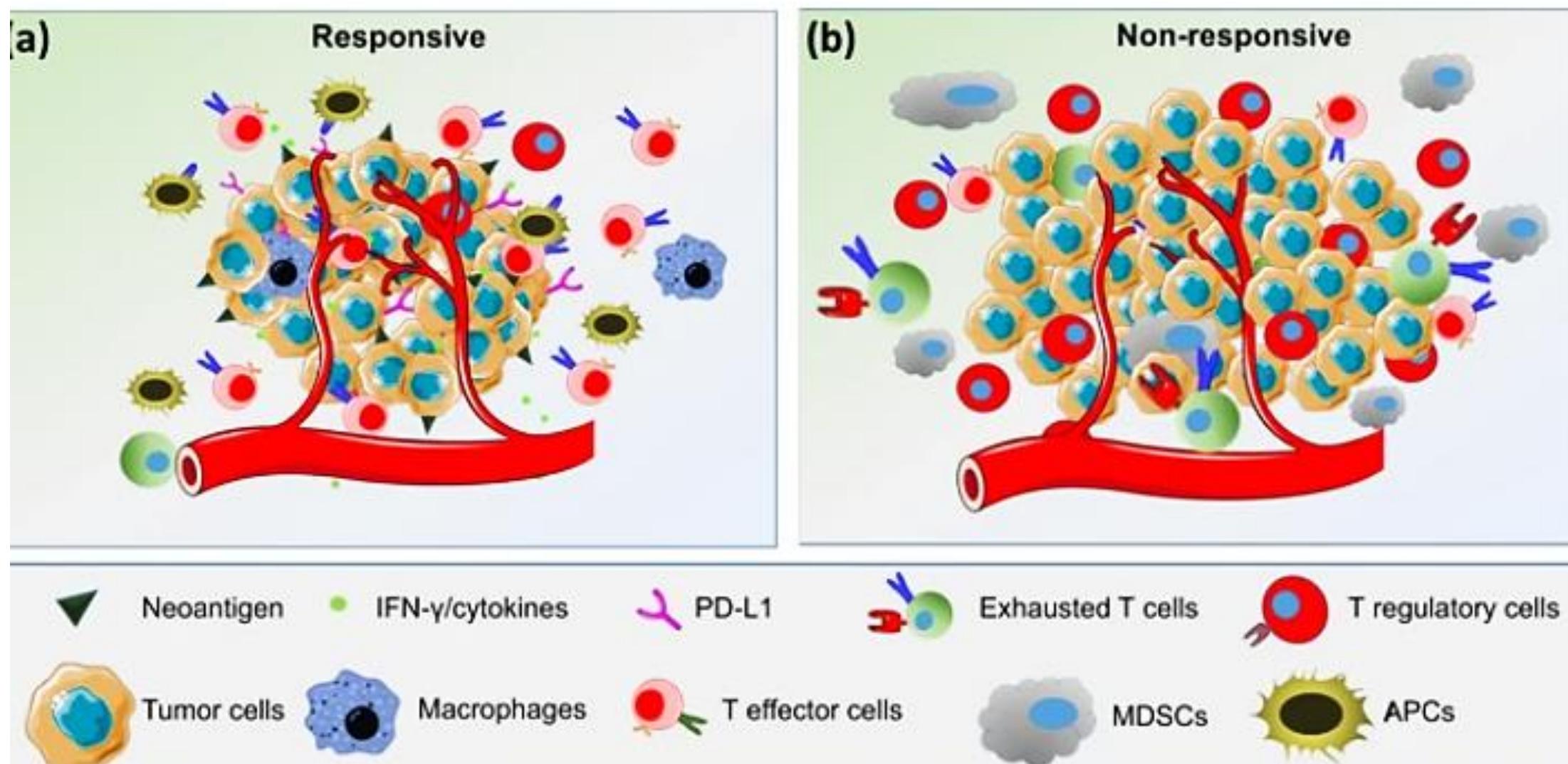


Early alterations in stem-like/marrow-resident T cells and innate and myeloid cells in preneoplastic gammopathy

Jithendra Kini Bailur,¹ Samuel S. McCachren,^{1,2} Deon B. Doxie,¹ Mahesh Shrestha,¹ Katherine Pendleton,¹ Ajay K. Nooka,^{1,3} Natalia Neparidze,⁴ Terri L. Parker,⁴ Noffar Bar,⁴ Jonathan L. Kaufman,^{1,3} Craig C. Hofmeister,^{1,3} Lawrence H. Boise,^{1,3} Sagar Lonial,^{1,3} Melissa L. Kemp,² Kavita M. Dhodapkar,^{3,5} and Madhav V. Dhodapkar^{1,3}

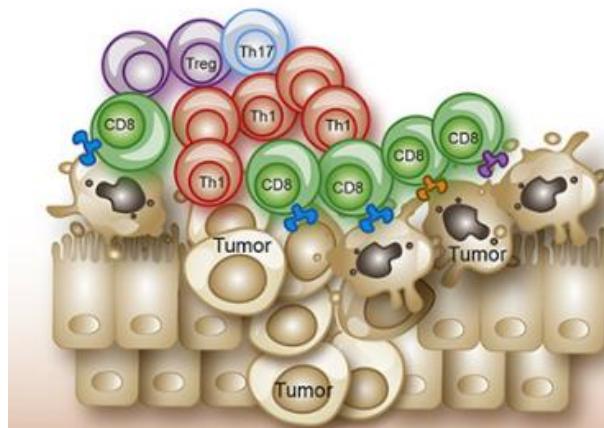
Early and complex alterations in the immune landscape in MGUS, including both innate and adaptive immune cells

Predictive biomarkers for response to ICP blockade



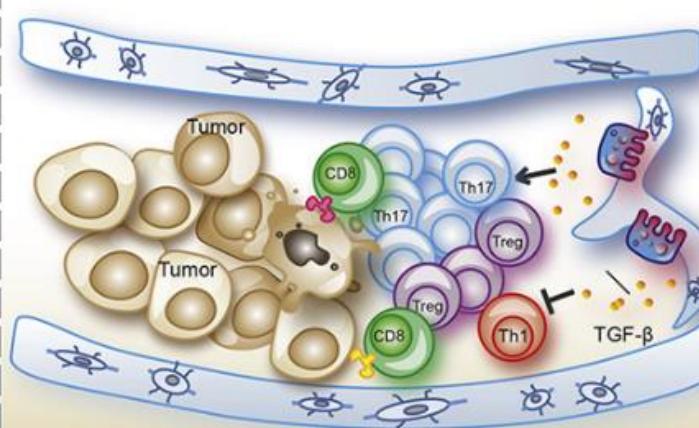
Tissue-Specific Checkpoint Immunotherapy Evasion

Checkpoint Therapy



Non-Osseous

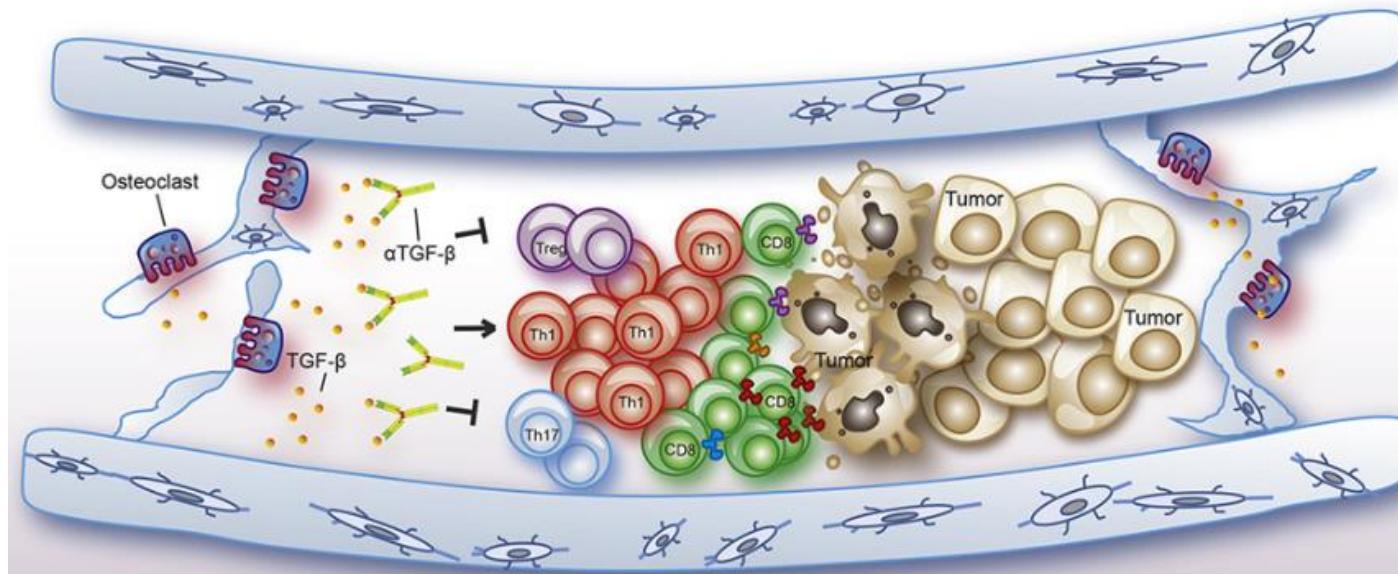
Checkpoint Therapy



Osseous

Checkpoint Therapy + α TGF- β

Osseous Environment

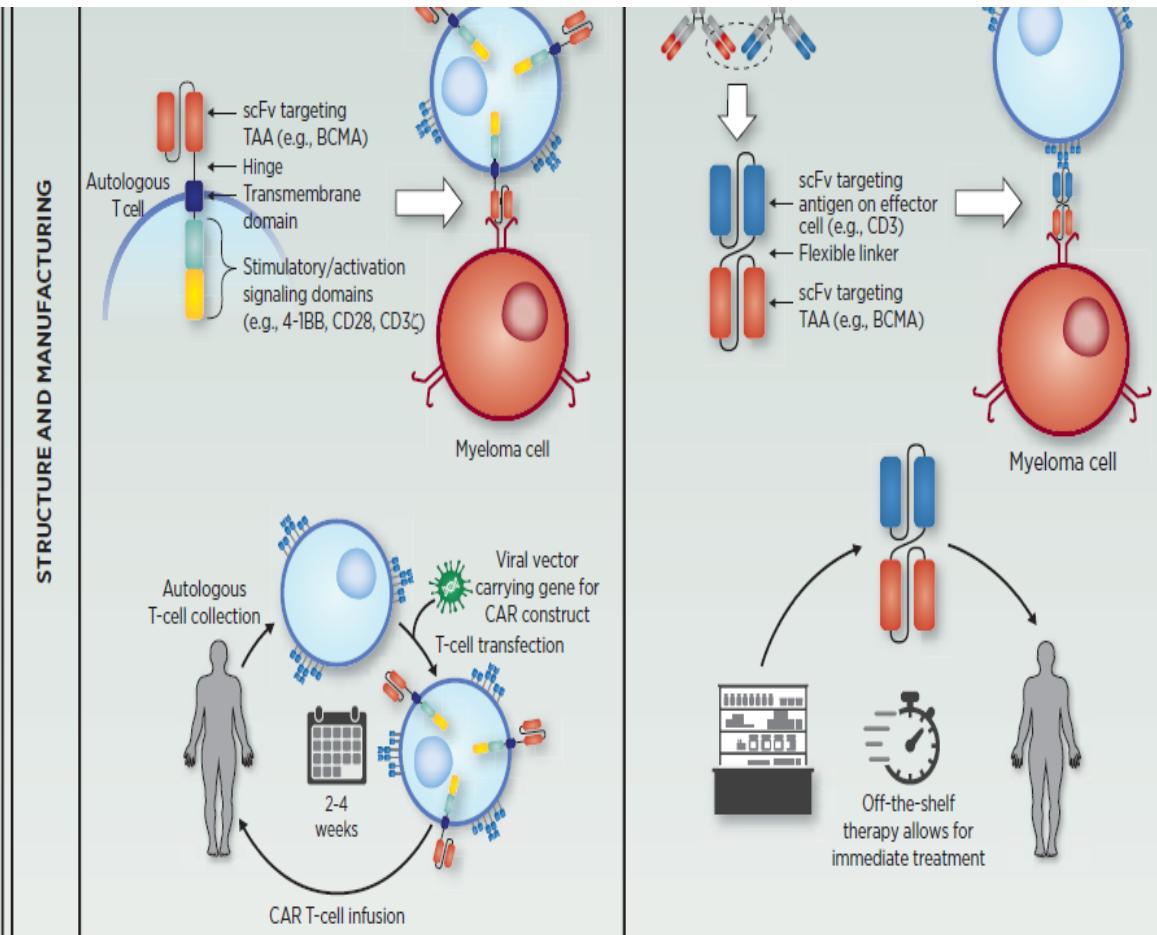


Disease progression

	MGUS	SMM	NDMM	SCT ^a	RRMM
Vaccines ^b	● DC DKK1 ^c NCT03591614 ^d ● Neoantigen NCT03631043	● PVX-410 NCT02886065	● SurVaxM NCT02334865	● PD-L1 peptide NCT03042793	● DC/MM fusion NCT00458653
	● PVX-410 NCT01718899	● GVAX NCT03376477		● Recombinant MAGE-A3 NCT01380145	● mRNA electro- porated DC ^e NCT01995708
	● PD-L1 peptide NCT03850522			● DC/MM fusion NCT02728102	● WT1 peptide (GPS) NCT01827137
Checkpoint Inhibitors ^f		● Pembrolizumab NCT02603887	● Pembrolizumab NCT02636010 ^f NCT02880228	● Pembrolizumab NCT02906332	● Pidilizumab NCT01067287 ^g
					● Pembrolizumab NCT03267888
					● Atezolizumab NCT02431208
					● Pembrolizumab NCT01953692
					● Pidilizumab NCT02077959
					● Nivolumab NCT02681302
					● Pembrolizumab NCT02362035
					● Nivolumab NCT0357952
					● Nivolumab NCT02726581
CART cells ^h			● CART-19/BCMA NCT03455972	● CAR-BCMA NCT02215967	● P-BCMA-101 NCT03288493
				● CART-BCMA NCT02546167	● BCMA CAR-T NCT03093168
				● bb2121 NCT02658929	● CTL019 CAR T NCT02135406
				● bb21217 NCT03274219	● NKG2D-CAR T NCT02203825
				● KITE-585 NCT03318861	● DesCartes-08 NCT03448978
				● CAR2 NCT03464916	● Kappa.CART NCT00881920
				● CART-138 NCT01886976	● BCMA CAR-T NCT03502577
				● JCARH125 NCT03430011	● NKR-2 NCT03018405
				● LCAR-B38M NCT03090659	● CART-138/BCMA/ 19/More NCT03196414
				● JNJ-68284528 NCT03548207	
				● AUTO2 CAR T NCT03287804	
BsAb/antibody constructs ⁱ				● Blinatumomab NCT03173430	● PF-06863135 NCT03269136
				● AMG 420 NCT02514239	● GBR-1342 NCT03309111
				● AMG 701 NCT03287908	● BFCR4350A NCT03275103
				● AMG 424 NCT03445663	● REGN-5458 NCT03761108

Study phase

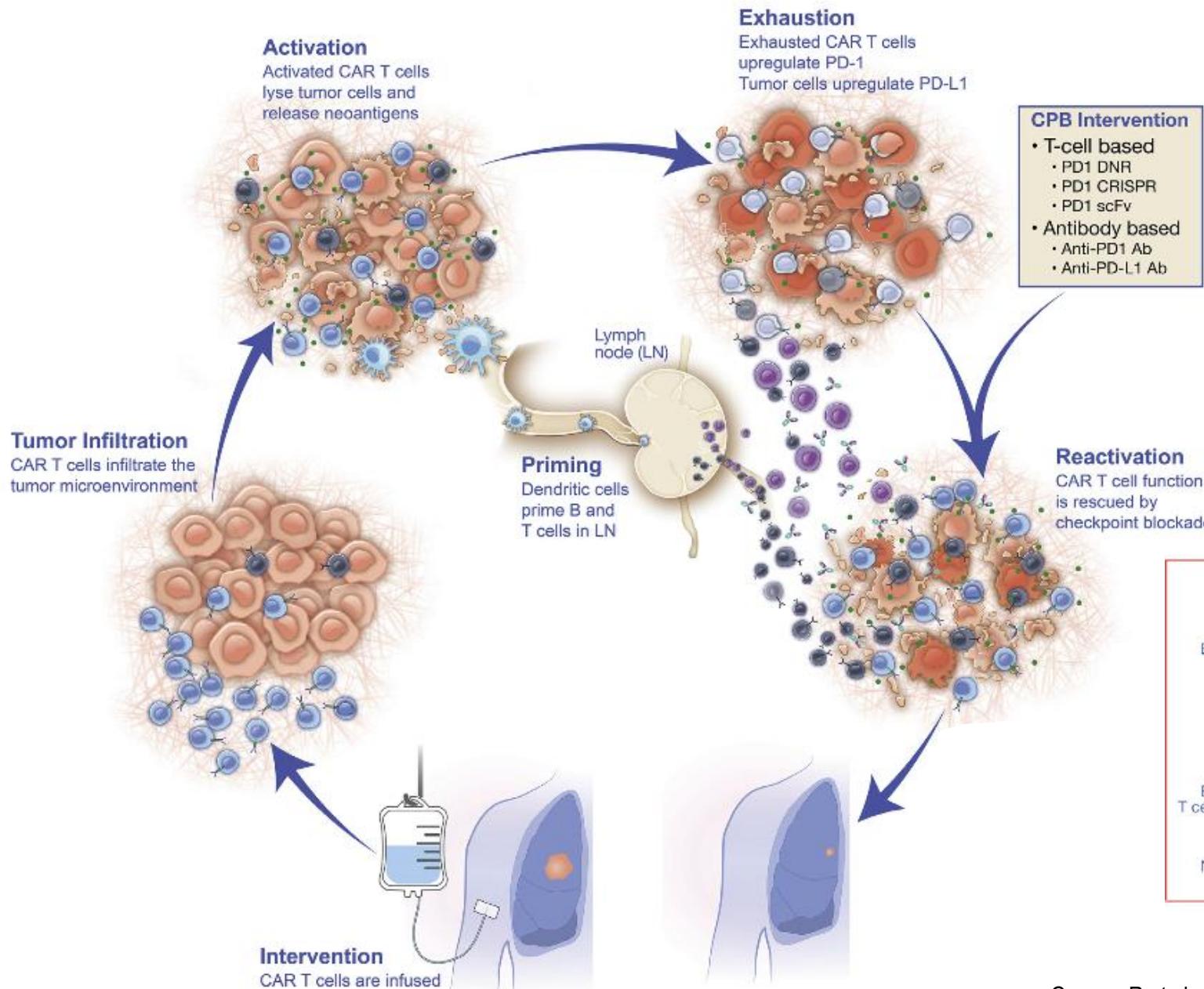
- Early phase 1
- Phase 1
- Phase 1/2
- Phase 2
- Phase 2/3
- Phase 3



CHALLENGES AND STRATEGIES FOR IMPROVEMENT

Challenge	Strategies for Improvement
Immunogenicity (e.g., HAMA)	<ul style="list-style-type: none"> Fully human product
Manufacturing time	<ul style="list-style-type: none"> Allogeneic product More rapid manufacturing protocols
Target antigen loss/antigen-negative relapse	<ul style="list-style-type: none"> Gamma-secretase inhibition for BCMA (e.g., NCT03502577) Dual antigen-targeting product
Persistence	<ul style="list-style-type: none"> MIL-based product Preferential transduction of T_{CM} and T_{SCM} cell PI3K inhibitor during manufacturing Product with defined CD4:CD8 ratio
Challenge	Strategies for Improvement
Immunogenicity (e.g., HAMA)	<ul style="list-style-type: none"> Humanize mAbs Generate mAbs from phage display libraries based on human sequence
Target antigen loss/antigen-negative relapse	<ul style="list-style-type: none"> Infusion of 2 bsAbs/antibody constructs targeting separate TAAs
Frequency of administration	<ul style="list-style-type: none"> Extended half-life product
Safety (CRS, neurotoxicity, infections)	<ul style="list-style-type: none"> Tocilizumab to treat or prevent therapy-associated CRS Consensus grading criteria and management algorithms for CRS and neurologic toxicity

Rescue of CAR-T cell exhaustion with ICP blockade



Strategies to combine CAR-T cells and ICP blockade

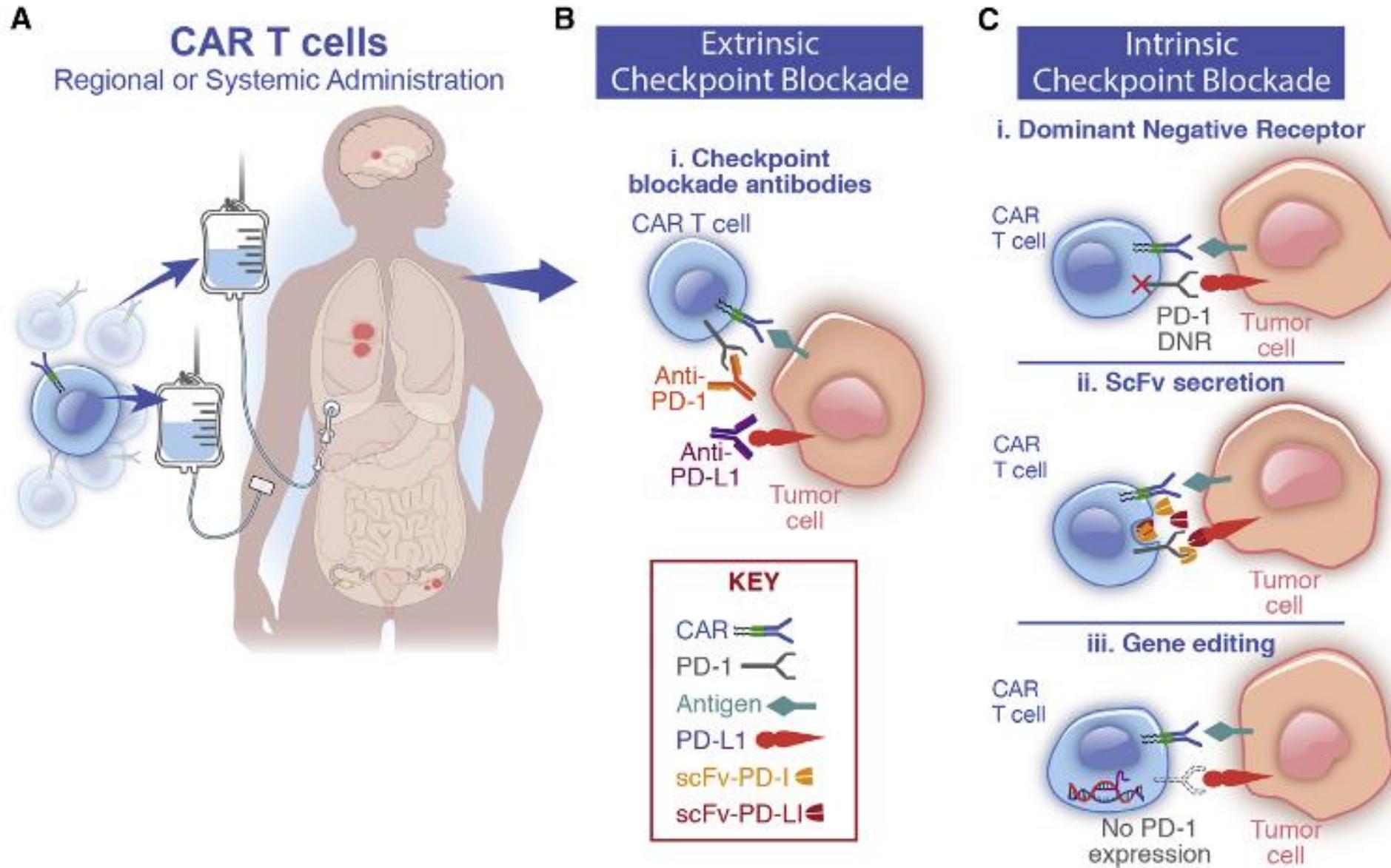


Table 1. Clinical Trials Exploring Combination Therapy with CAR T Cells and Checkpoint Blockade

Trial	Launch	Phase	Center(s)	CPB Agent	CAR Target/Design	Cancer Diagnosis
NCT00586391	2009	I	Baylor	ipilimumab	CD19/CD19CAR-28-zeta T cells	B cell lymphoma, chronic lymphocytic leukemia, acute lymphocytic leukemia
NCT01822652	2013	I	Baylor	pembrolizumab	GD2/iC9-GD2-CD28-OX40 (iC9-GD2) T cells	neuroblastoma
NCT02650999	2016	I/II	University of Pennsylvania	pembrolizumab	anti-CD19 CARs	CD19 ⁺ diffuse large B cell lymphoma, follicular lymphoma, mantle cell lymphoma
NCT02706405	2016	I	Fred Hutchinson	durvalumab	autologous anti-CD19CAR-4-1BB-CD3ζ-EGFR ^t -expressing CD4 ⁺ /CD8 ⁺ central memory T lymphocytes JCAR014	diffuse large B cell lymphoma
NCT02926833	2016	I/II	City of Hope, Stanford, Moffitt, Dana Farber, MD Anderson	atezolizumab	CD19/KTE-C19	diffuse large B cell lymphoma
NCT03310619	2017	I/II	City of Hope, Northwestern University, Massachusetts General, University of Nebraska, University of Pennsylvania, MD Anderson	durvalumab	JCAR017	lymphoma, non-Hodgkin lymphoma, diffuse large B cell lymphoma, follicular lymphoma
NCT03726515	2018	I	University of Pennsylvania	pembrolizumab	CART-EGFRvIII T cells	glioblastoma

Immune stimulation

- Vaccines
- CAR-T
- NK/NKT/γδ

Immune modulators

- macrophage inhibitors
- immune-stimulatory agents

Epigenetic modifications

- HMA
- HiDAC

metabolic modulators

- IDO inhibitors
- A2AR inhibitors
- anti-CD73

ICP blockade

Targeted Therapy:

- BRAF+MEK inhibitors
- VEGF inhibitors
- BRAF+MEK inhibitors
- EGFR inhibitors
- VEGF inhibitors
- PI3K delta

Chemotherapy
(immunogenic cell death)

Radiotherapy
(abscopal effect)

Credits

Laboratory of Blood Tumor Immunology, CeRMS

Barbara Castella (Senior Lab Investigator)

Myriam Foglietta

Ezio Tripoli

Claudia Giannotta

Assunta Melaccio



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Claudia Castellino

Myriam Foglietta

Mariella Grasso

Daniele Mattei

Nicola Mordini

Davide Rapezzi

Roberto Sorasio



Department of Oncology, Turin, Italy

(Prof.ssa Chiara Riganti)

Joanna Kopecka

Laboratory of Immunogenetics, Turin

(Prof.ssa Ada Funaro)

Angelo Corso Faini

Yulia Yakymiv

Lab of Angiogenesis and Immunology

Bari (Prof. Angelo Vacca)

Roberto Ria

Assunta Melaccio



2ST CUNEO CITY IMMUNOTHERAPY CONFERENCE (CCITC)
IMMUNOTHERAPY IN HEMATOLOGICAL MALIGNANCIES 2020
JUNE 18-20, 2020

ORGANIZED BY MASSAIA M, SC EMATOLOGIA AO S.CROCE E CARLE, CUNEO, ITALY
& CENTRO INTERDIPARTIMENTALE DI RICERCA IN BIOLOGIA MOLECOLARE (CIRBM), TORINO, ITALY