

Highlights from IMW 2019

19-20 novembre 2019
Bologna
Royal Hotel Carlton

Aldo M. Roccaro, MD, PhD
Come e quando
valutare l'MRD
nel sangue periferico

Coordinatore Scientifico
Michele CAVO

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Mario BOCCADORO
Michele CAVO
Maria Teresa PETRUCCI



Conflict of Interest Disclosure:

Aldo M Roccaro

Advisory board:

Celgene

Janssen

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Fondazione AIRC

European Hematology Association

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AstraZeneca

Rational for testing MRD in MM using peripheral blood samples

- ✓ Avoid serial assessments requiring repeated BM aspirate sampling
 - ✓ Negative MRD may be obtained as a result of dilution of BM with blood and/or the patchy distribution of transformed plasma cells
-

Are malignant plasma cells detectable within a peripheral blood specimen?

Flow cytometry
NGS
Mass spectrometry

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Detection of Myeloma Cells in the Peripheral Blood by Flow Cytometry

Thomas E. Witzig, Teresa K. Kimlinger, Gregory J. Ahmann, Jerry A. Katzmann, and Philip R. Greipp

Division of Internal Medicine and Hematology (T.E.W., P.R.G.) and Department of Laboratory Medicine and Pathology (T.K.K., G.J.A., J.A.K.), Mayo Clinic and Mayo Foundation, Rochester, Minnesota

Cytometry (Communications in Clinical Cytometry) 26:113–120 (1996)

AIM: to identify circulating plasma cells from whole blood specimens, using

- ✓ two-color FC (CD38; CD45)
- ✓ three-color FC (CD38; CD45; κ ; λ)

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Flow cytometry of mononuclear cells from a patient with IF microscopy- documented monoclonal λ plasma cells

$CD38^{bright}/CD45^{neg}/\lambda$

$CD38^{bright}/CD45^{dim}/\lambda$

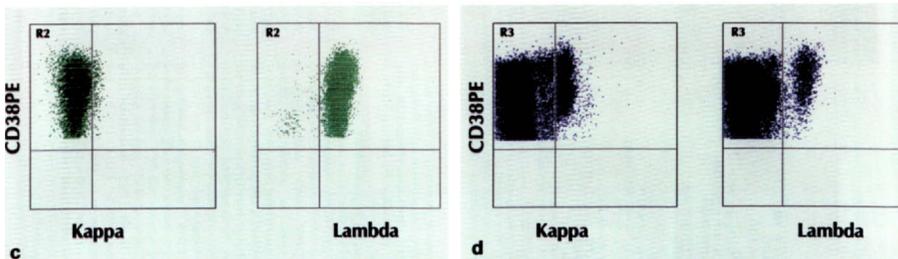
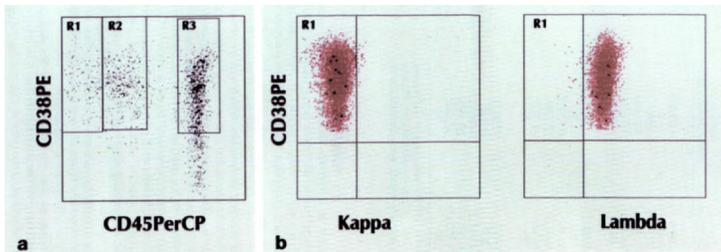


FIG. 4. Flow cytometry dot-plots of the mononuclear cells from a patient with documented monoclonal λ plasma cells by immunofluorescence microscopy. **a:** CD45PerCP vs. CD38PE. Gate 1 contains CD38^{bright}CD45⁻ cells; Gate 2, CD38^{bright}CD45^{dim} cells; and Gate 3, CD38^{bright}CD45⁺ cells. **b, c,** and **d:** The kappa/lambda analysis on the cells in gates R1, R2, and R3, respectively. Monoclonal lambda cells are found predominantly in the CD38^{bright}CD45⁻ and CD38^{bright}CD45^{dim} cell populations.

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Out of the IF microscopy positive cases

two-color: detection 94%

Three-color: detection 77%

Whole Blood Flow Cytometry

Ab anti-CD38 and –CD45

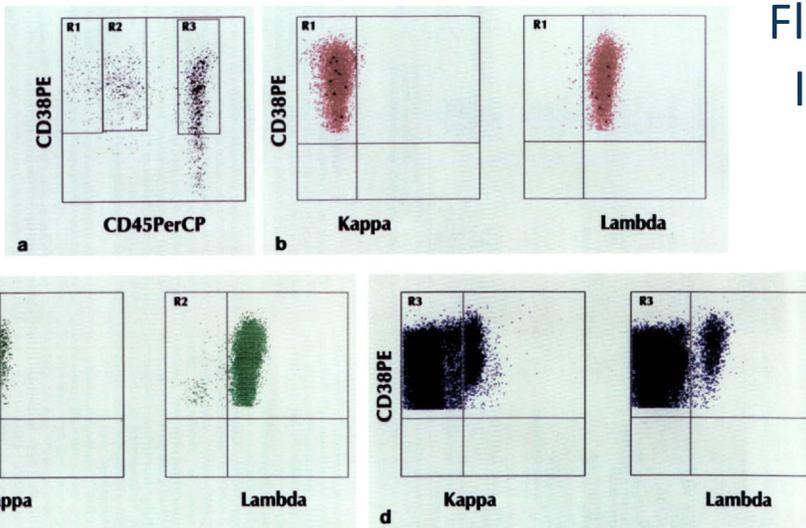


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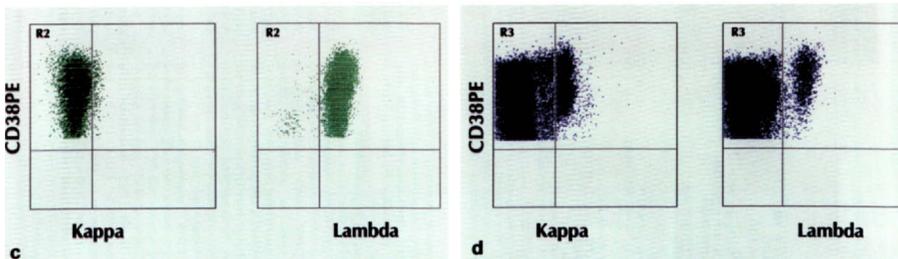
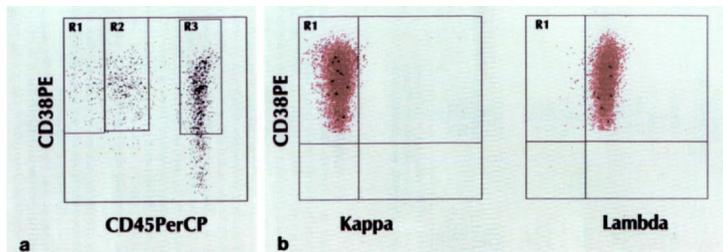


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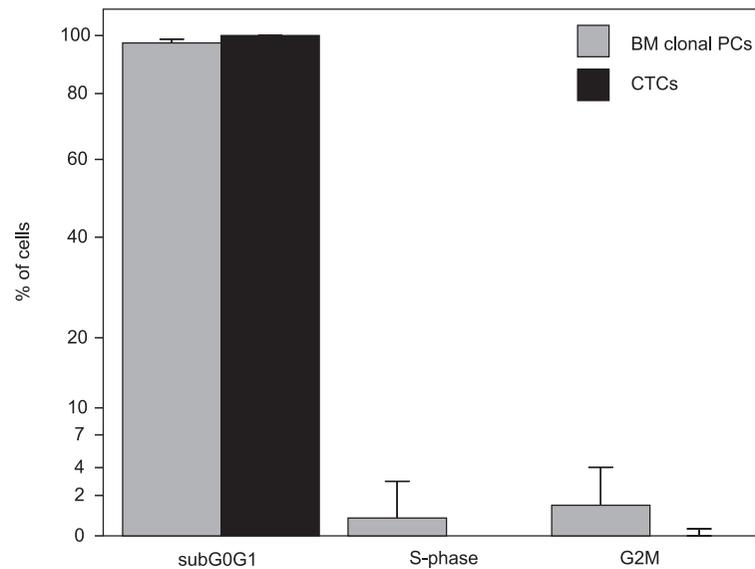
Whole Blood Flow Cytometry

Ab anti-CD38 and -CD45

Practical and reliable method to detect and quantify circulating malignant myeloma plasma cells

Witzig et al, Cytometry, 1996

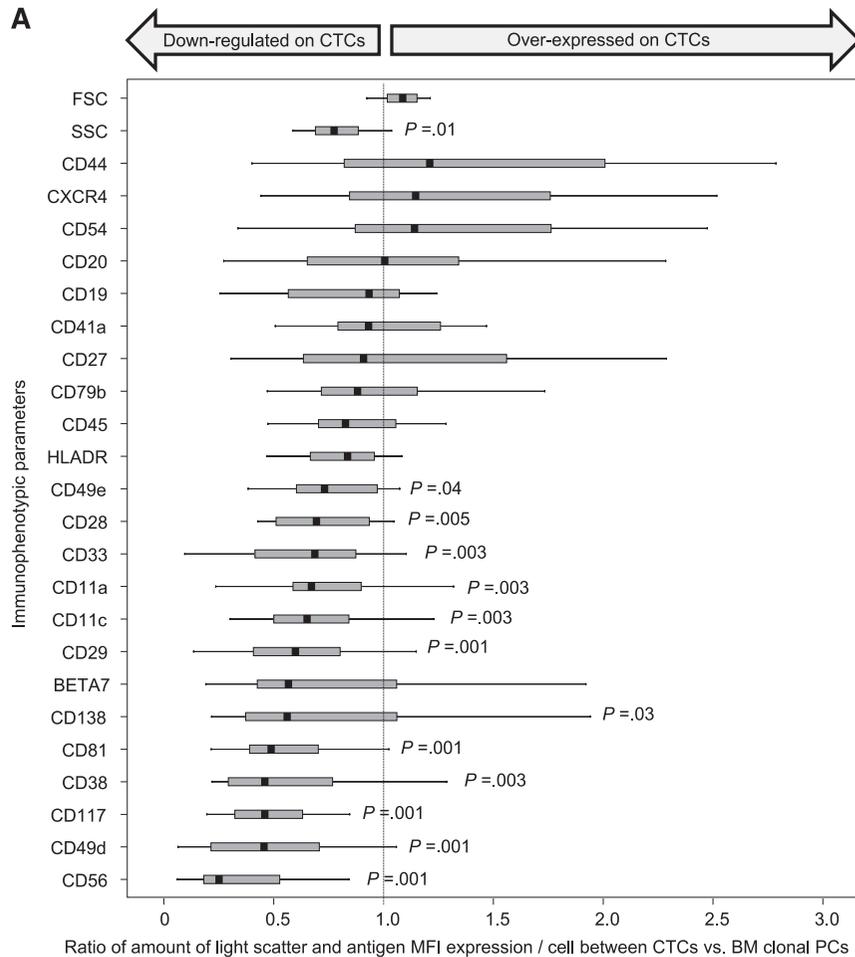
CTCs represent a unique subset of the whole BM clonal plasma cells compartment



circulating neoplastic PCs vs matched BM

- ✓ mostly quiescent: arrested in the subG0-G1
- lower S-phase
- lower G2/M

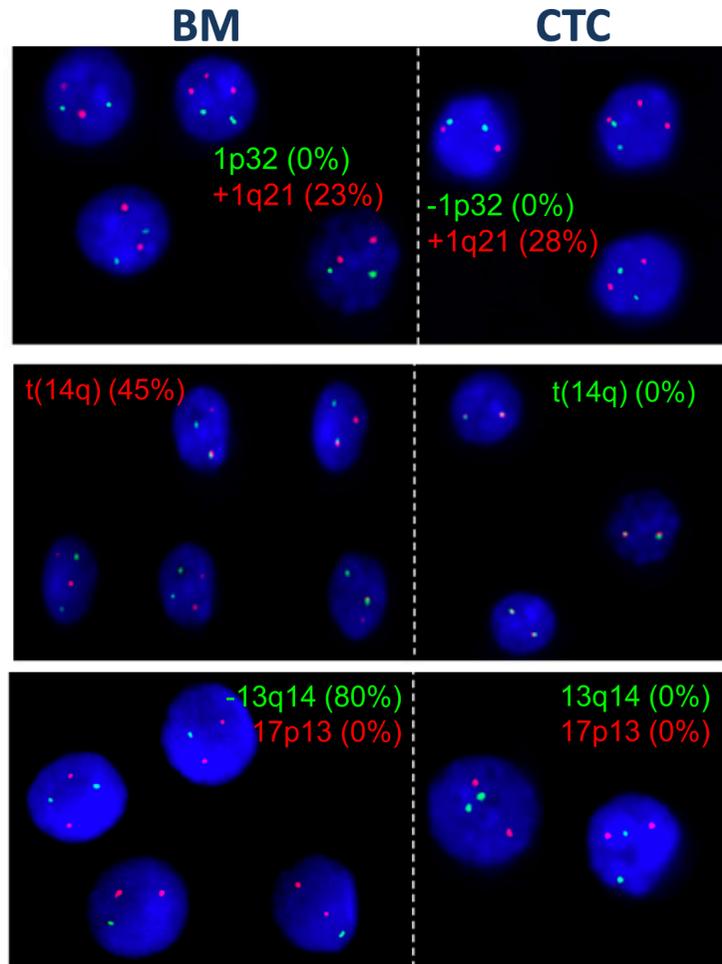
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downregulation ($P < .05$) of:

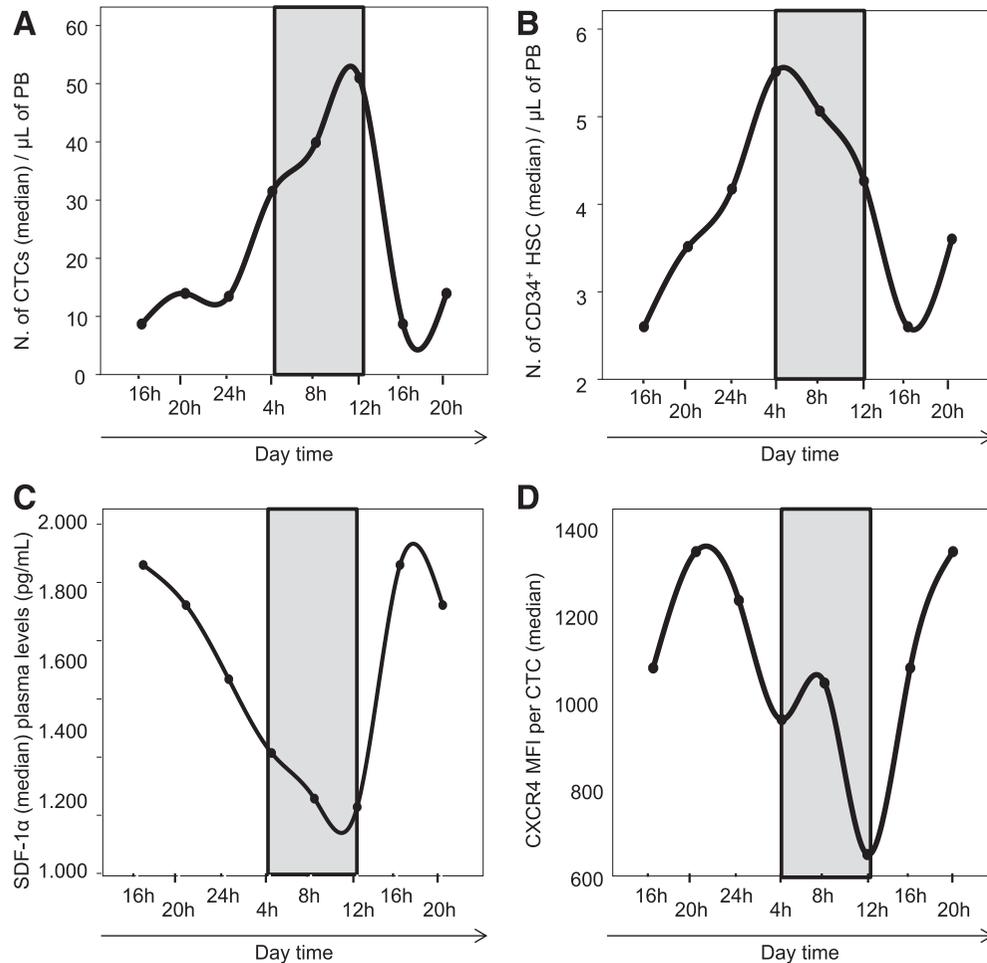
- ✓ integrins
(CD11a/CD11c/CD29/CD49d/CD49e)
- ✓ adhesion (CD33/CD56/CD117/CD138)
- ✓ activation molecules (CD28/CD38/CD81)

CTCs represent a unique subset of the whole BM clonal plasma cells compartment



circulating neoplastic PCs vs matched BM
✓ different cytogenetic profile

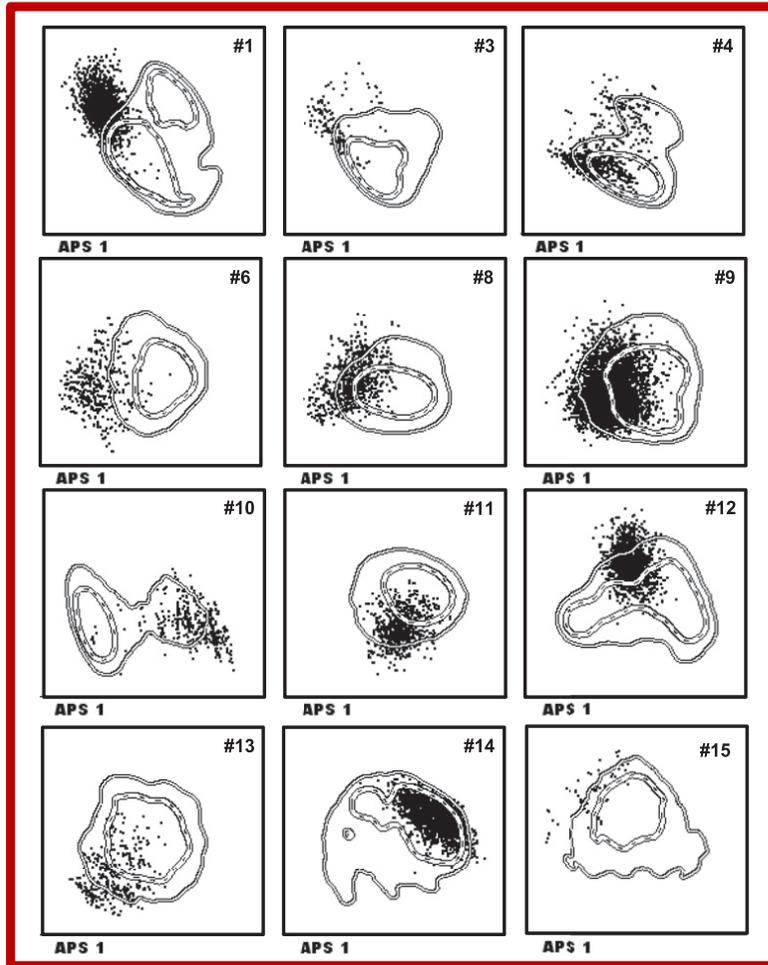
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CTCs: circadian distribution

- ✓ similar pattern to CD34 cells
- ✓ opposite to SDF-1 plasma levels and CXCR4 surface expression on clonal PCs
- ✓ CTCs may egress to PB to colonize/metastasize other sites in the BM during the patients' resting period

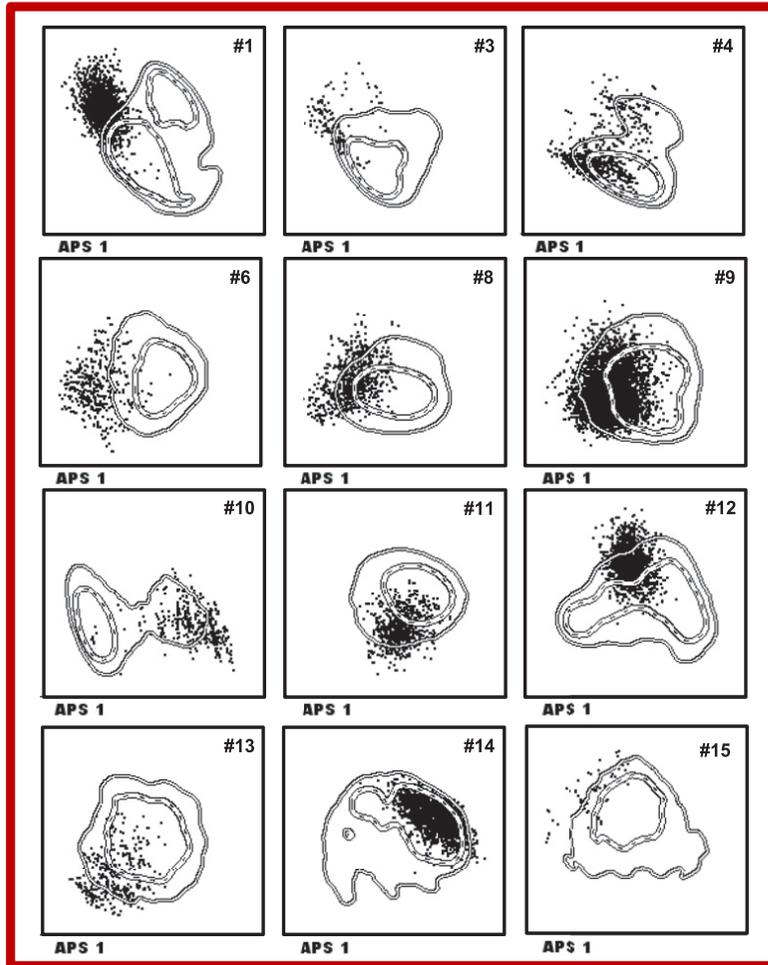
CTCs represent a unique subset of the whole BM clonal PC compartment



✓ CTCs clustered in a uniquely restricted area of that occupied by BM clonal PCs: 12/15

iPEP of BM clonal PCs is represented by 1 and 2 SD lines
paired CTCs are represented by black dots.

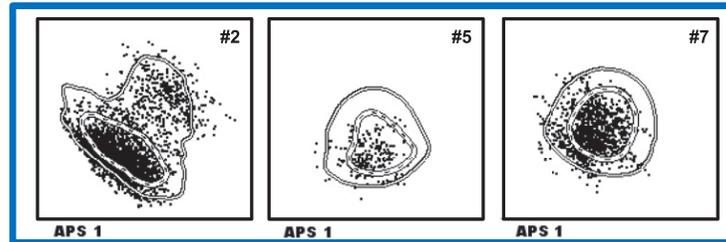
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compartment

✓ CTCs clustered in a uniquely restricted area of that occupied by BM clonal PCs: 12/15

✓ CTCs were spread throughout the whole BM clonal PC compartment: 3/15

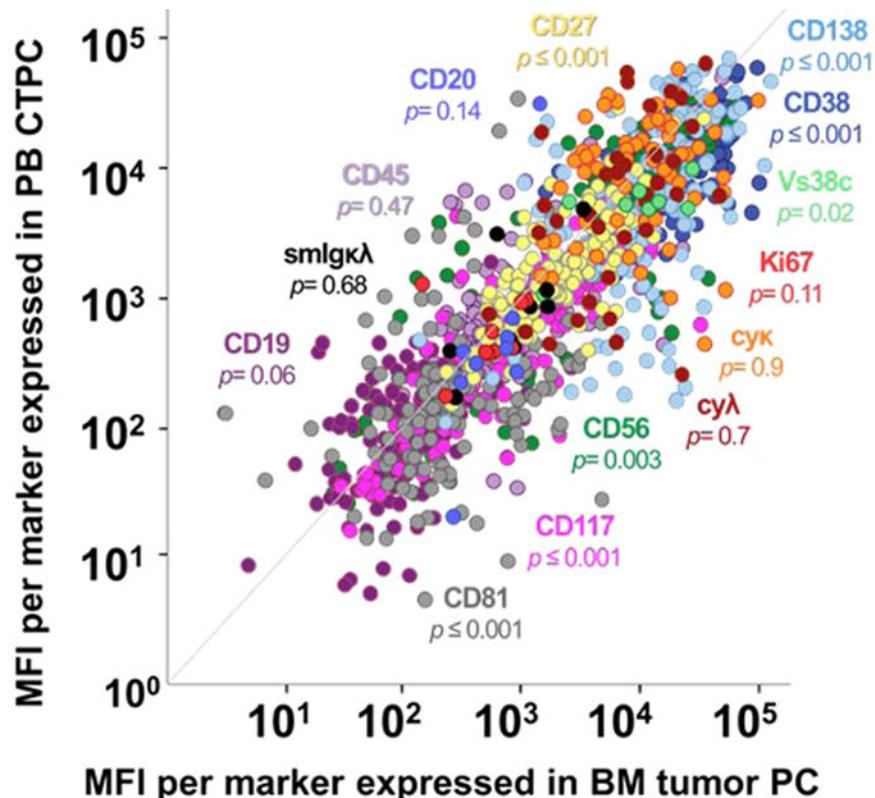


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**CTCs represent a unique subset
of the whole BM clonal plasma cells compartment**

**Detailed characterization of multiple myeloma circulating tumor
cells shows unique phenotypic, cytogenetic, functional, and
circadian distribution profile**

Correlation between the immunophenotype of tumor PCs in paired PB and BM samples from newly diagnosed MM



PB-CTPCs vs BM-TPCs: similar immunophenotypic profile

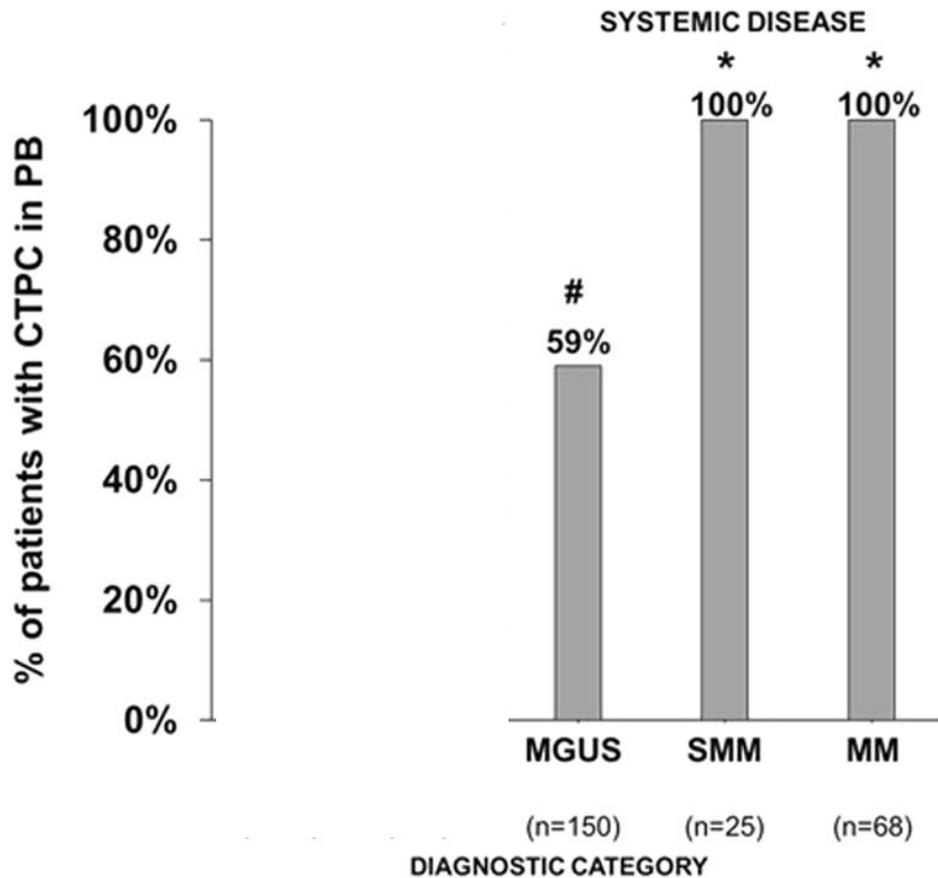
PB-CTPCs significantly lower ($p < 0.05$) of CD38, CD138, CD81, CD56, CD27, and Vs38c maturation-associated markers, CD117

more immature and less proliferative immunophenotype for paired PB vs. BM

PB-CTPCs: peripheral blood-circulating tumor plasma cells
BM-TPCs: bone marrow-tumor plasma cells

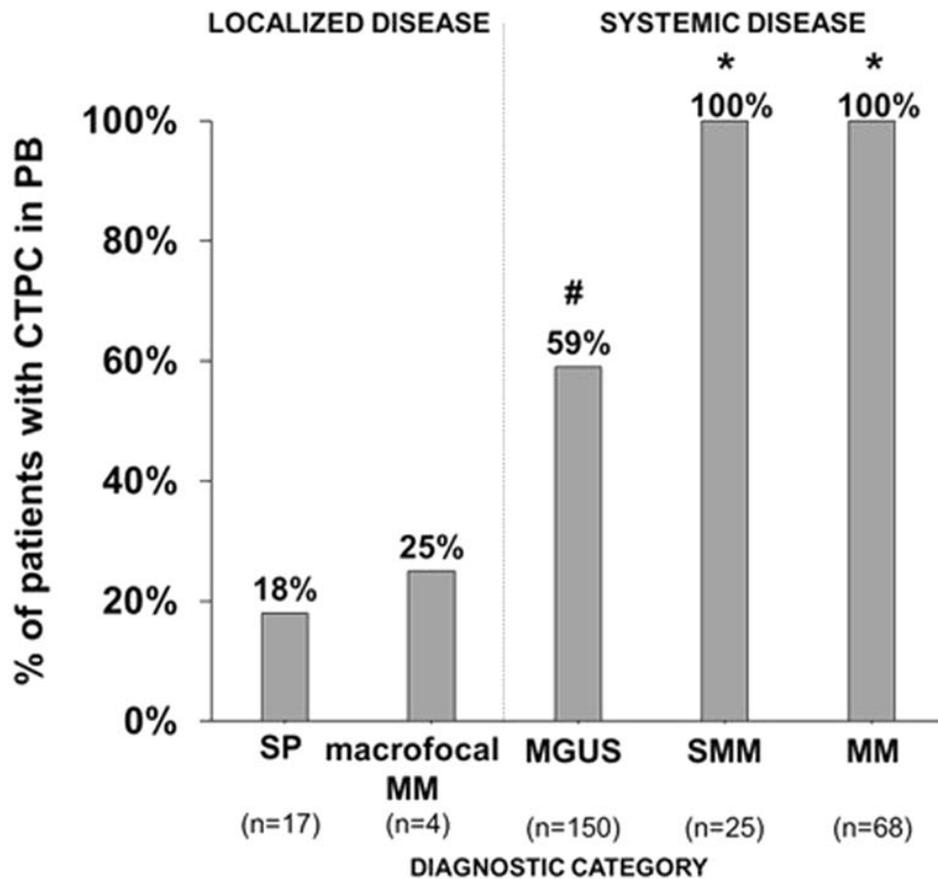
Flores-Montero J, et al. Leukemia 2017

Detection of tumor plasma cells within the peripheral blood of patients with plasma cell neoplasias - NGF



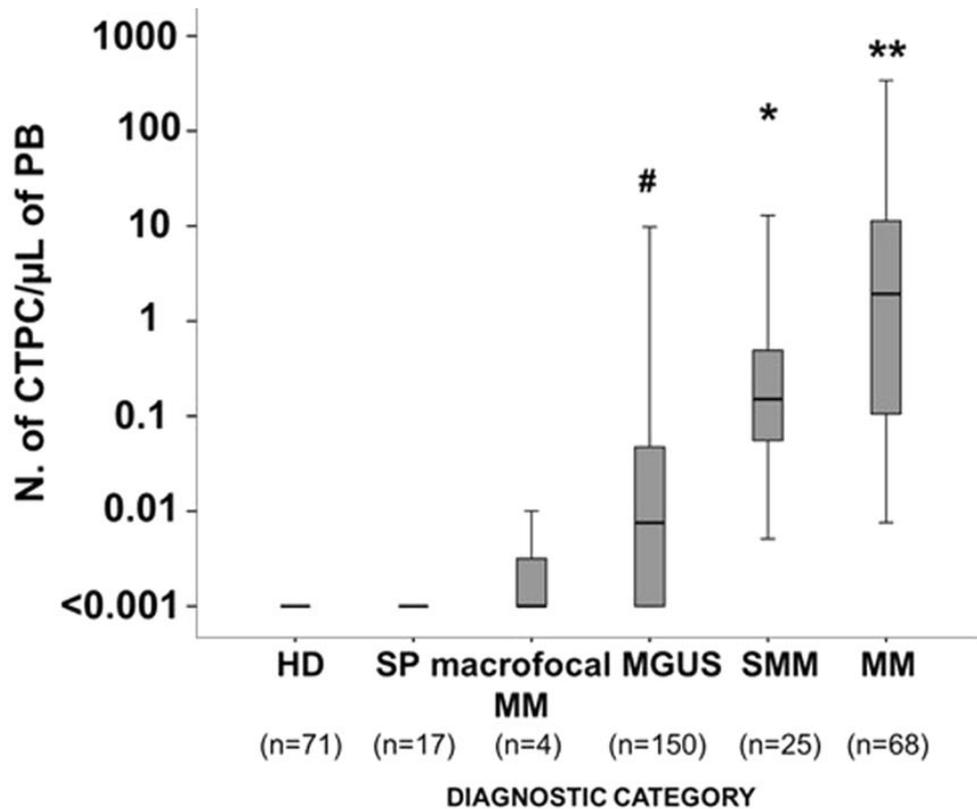
Presence of CTPC in PB is associated with systemic disease (i.e., MGUS, SMM and MM) vs localized disease (i.e., macrofocal MM, solitary plasmacytoma)

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Overall number of PB CTPC as assessed by NGF also increased progressively from SP and macrofocal MM to MGUS, SMM, and MM

Detection of tumor plasma cells within the peripheral blood of patients with plasma cell neoplasias - NGF

All treated MM patients who showed CTPC after therapy always showed MRD + of paired BM samples (data not shown)

persistence/presence of CTPC in MM patients who had undergone therapy, might be used as a surrogate marker of BM MRD-positivity

Preliminary results

Further validation needed

Are malignant plasma cells detectable within a peripheral blood specimen?

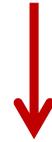
Flow cytometry

NGS

Mass spectrometry

Detection of tumor plasma cells within the peripheral blood of patients with plasma cell neoplasias evaluating ctDNA

- ✓ Circulating tumor DNA: promising non-invasive tool, for monitoring response to therapy
- ✓ Clonotypic V(D)J rearrangement: monitoring MM ctDNA after treatment initiation
Non-responders/progressors: detectable ctDNA at times of high tumor burden compared with less than half of responders



**MRD by NGS Clonotypic V(D)J rearrangement:
peripheral blood vs matches BM samples**

Myeloma MRD by deep sequencing from circulating tumor DNA does not correlate with results obtained in the bone marrow

Table 1. Comparison of MRD status obtained in plasma and in bone marrow

	Plasma MRD positive	Plasma MRD negative	Total
Bone marrow MRD positive	8	18	26
Bone marrow MRD negative	1	10	11
Total	9 (PPV _{plasma} = 89%)	28 (NPV _{plasma} = 36%)	37

NPV_{plasma}, negative predictive value of MRD assessed from plasma sample; PPV_{plasma}, positive predictive value of MRD assessed from plasma sample.

- ✓ At the time of MRD (n. 37): 49% (18/37) consistency between paired plasma and BM

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- ✓ Most frequent discrepancy: undetectable MRD in plasma, which was BM positive (18/26)
- ✓ MRD assessment of ctDNA: only 36% negative predictive value; 89% positive predictive value
 - ✓ No quantitative correlation between plasma and BM was found, including when MRD was positive in both samples

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Only 1 discrepant case in which MRD was plasma-positive and BM-negative

A) Extramedullary (EM) relapse → PET/CT negative for EM relapse (at the time of MRD)

B) False-negative result in BM → Not PB-diluted
Patchy nature of the disease within the BM
Possibly, additional BM sample analysis may have revealed MRD positivity within the BM

Myeloma MRD by deep sequencing from circulating tumor DNA does not correlate with results obtained in the bone marrow

Only 39% of patients with less than a VGPR displayed detectable ctDNA

ctDNA was undetectable in 69% of patients with MRD detected within the BM



ctDNA may not serve as a sufficient analyte for MRD monitoring

Are malignant plasma cells detectable within a peripheral blood specimen?

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NGS

Mass spectrometry

Peripheral blood approached by using mass spectrometry

mass spectrometry

targets the (M- protein) similar to conventional electrophoresis

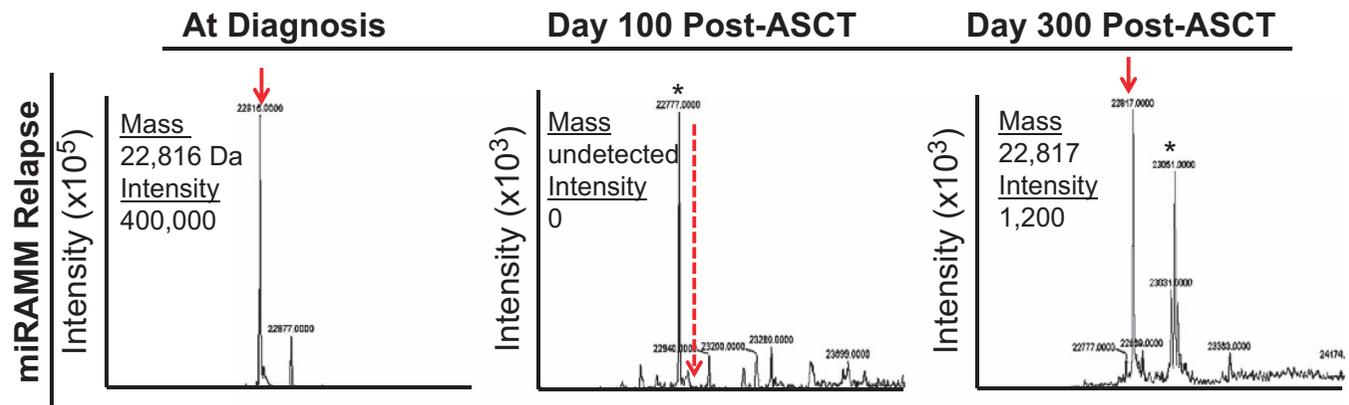
looks for the unique mass of the unique amino acid sequencing on each monoclonal Ig



much higher sensitivity

Peripheral blood approached by using mass spectrometry

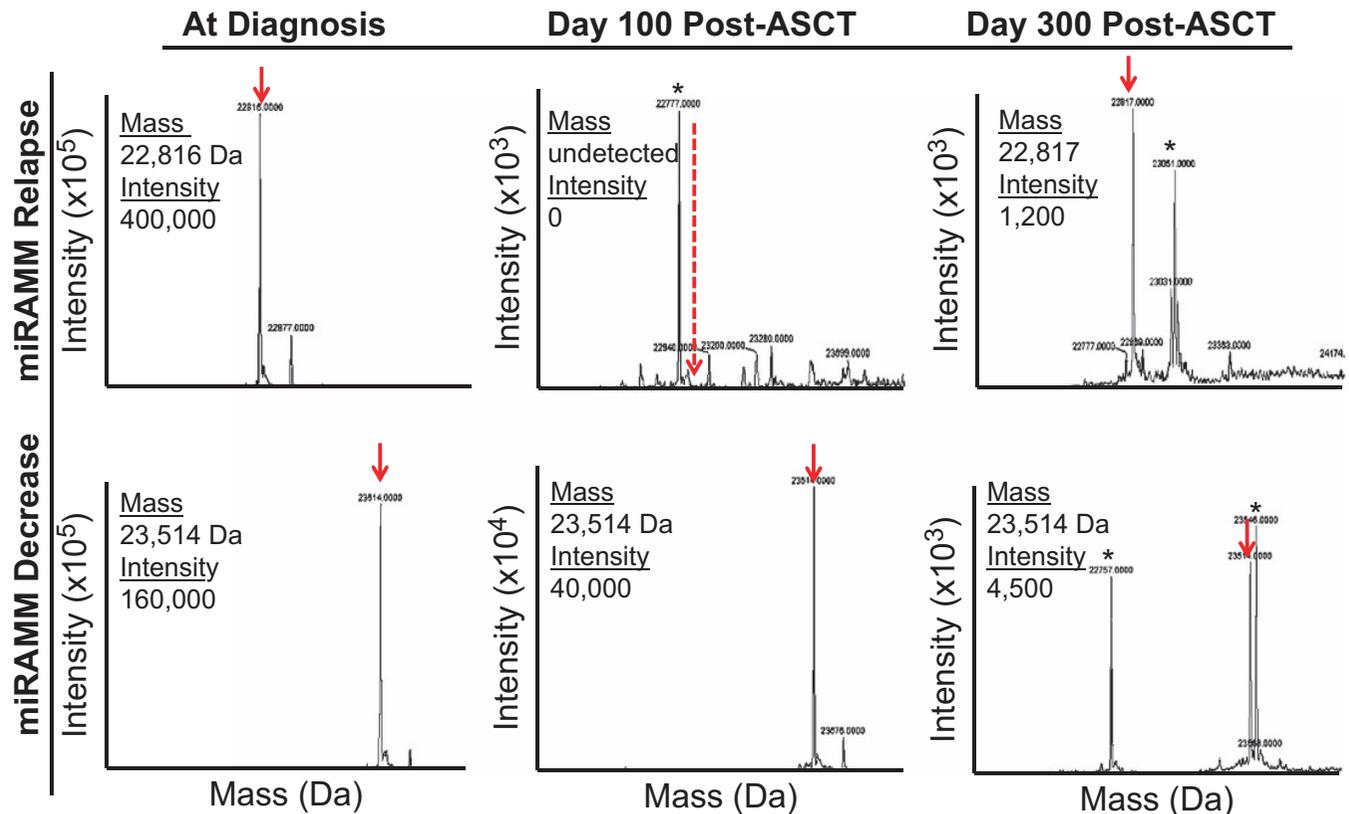
miRAMM: monoclonal immunoglobulin rapid accurate mass measurement
(sCR, post ASCT)



miRAMM relapsed

Peripheral blood approached by using mass spectrometry

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miRAMM relapsed

deepening miRAMM response

Peripheral blood approached by using mass spectrometry

Eveillard M, et al. ASCO 2019, #e19525

Matrix-Assisted Laser Desorption/Ionization - Time-Of-Flight mass spectrometer

MALDI-TOF in PB vs flow cytometry BM-based

MRD results were concordant for 44/71 (62%)
patients (8+/, 36 -/- respectively)
27 were discordant (10 +/-, 17-/+)

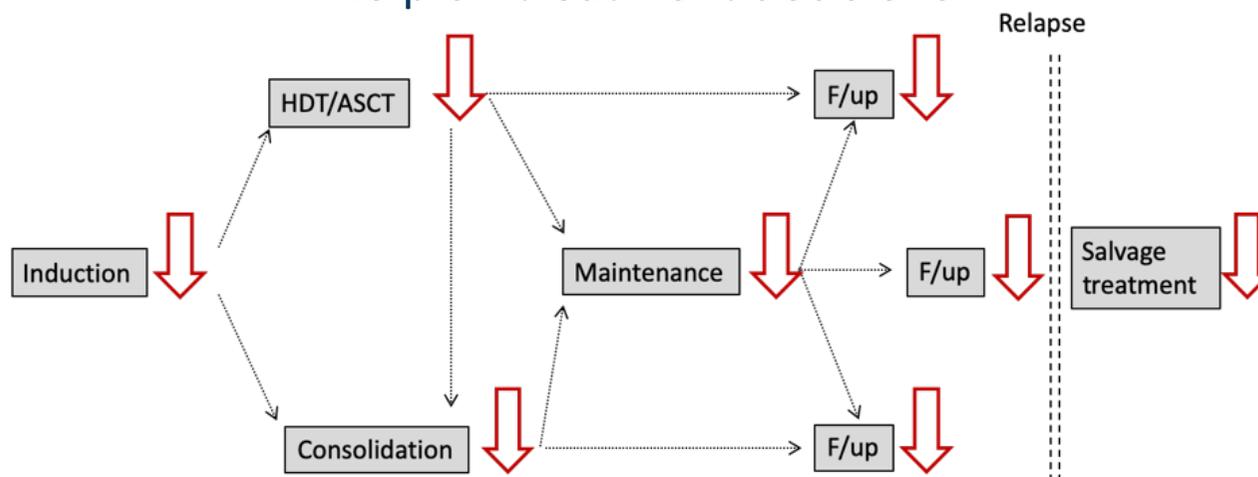
When to evaluate MRD?

With courtesy and permission of Dr. B. Paiva

IMW 2019, Boston, MA, USA

MRD evaluation is prognostic at any time point. Consider evaluating to:

- ✓ Define quality of CR
- ✓ Evaluate efficacy of subsequent treatment after CR (eg. Consolidation)
 - ✓ Identify high risk patients
 - ✓ Help on treatment decisions

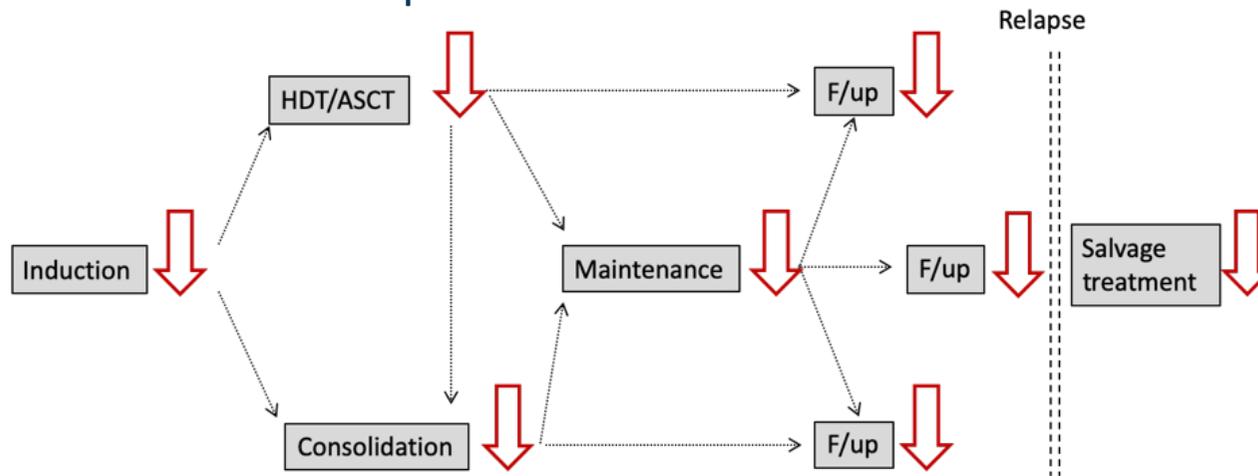


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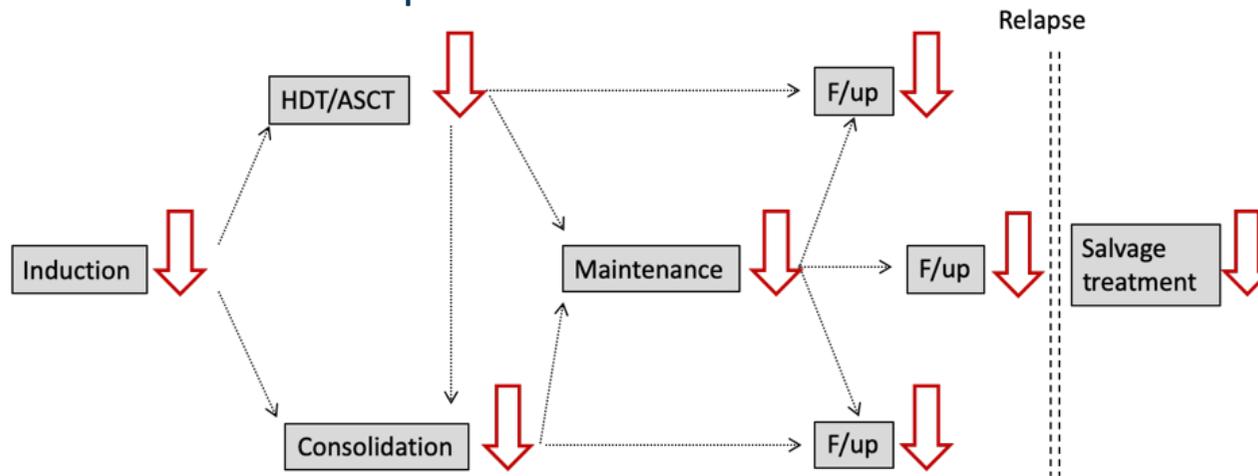
Bone marrow-based MRD

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Peripheral blood-based MRD: TBD

Bone marrow-based MRD

Take-home Points

- ✓ Existing recommendations for MRD testing (i.e., to be performed for VGPR in addition to CR) are all related to the BM-based evaluation.
 - ✓ One-time, blind BM aspiration biopsy is not able to provide information of the cancer on its entirety and can introduce bias and false negatives. A peripheral blood approach could potentially overcome these problems.
 - ✓ ctDNA may not serve as a sufficient analyte for MRD monitoring.
 - ✓ CTCs: possible source for monitoring MRD by NGF.
 - ✓ Mass-spectrometry-based assay are being taken into consideration.
-